

Figure 1

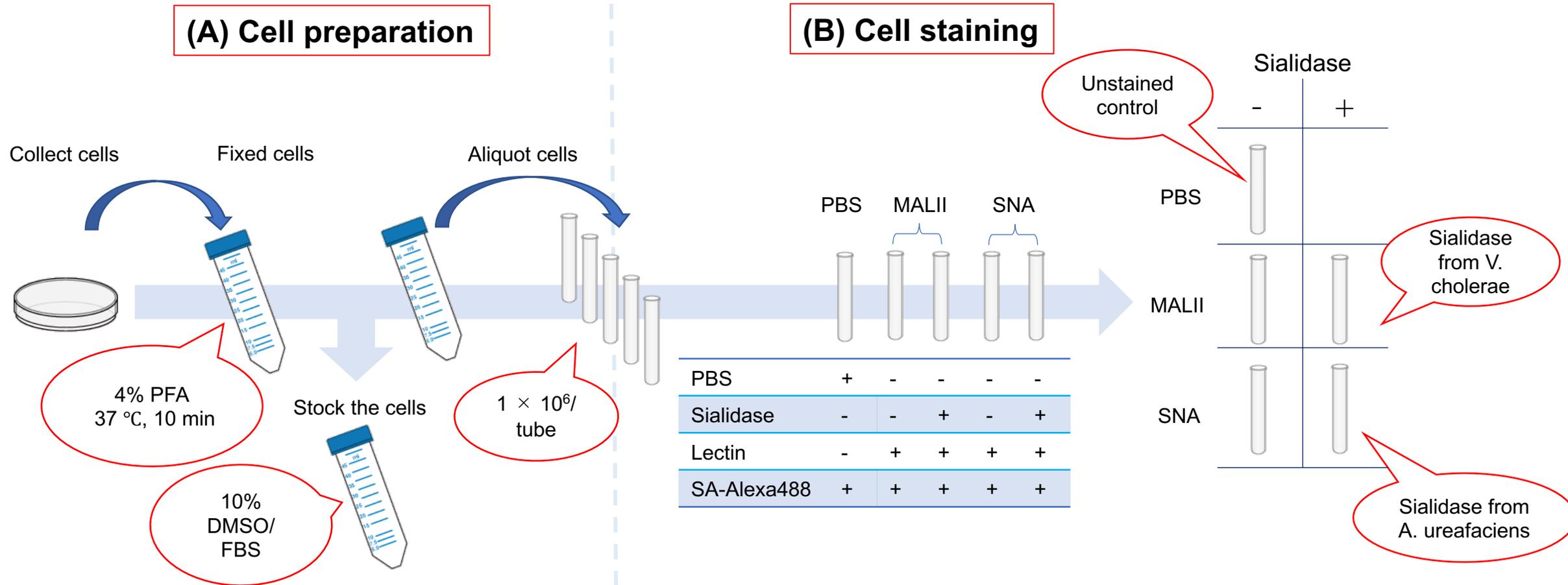


Figure 1. Overview of the procedure

(A) Cell preparation: cells are collected and fixed in 4%PFA. After fixing cells, the experiment could be suspended if necessary, and the cells could be stocked in 10% DMSO/ FBS. (B) Cell staining: the cells were divided into 5 tubes, and two out of 5 tubes are treated with sialidase for making the sialic acid-removing control (sialidase control). Then, biotinylated lectins are added two tubes including sialidase control tubes. PBS are added the remaining tube for control. After staining each sialic acid with the specific lectins, streptavidin-Alexa flour 488 is added into all tubes to detect the biotin.

Figure 2

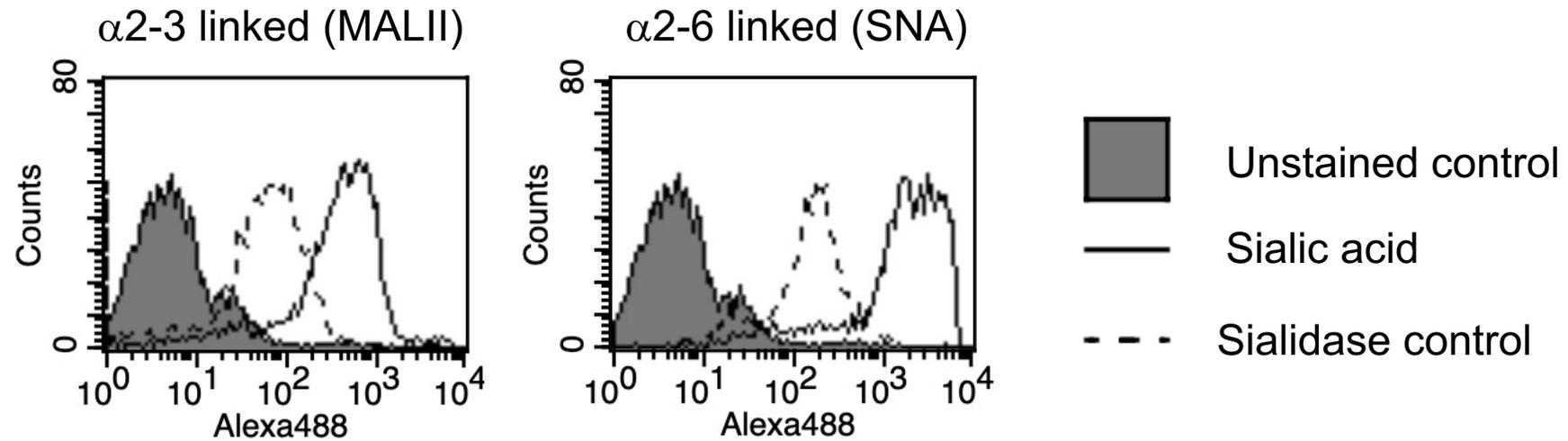


Figure 2. Sialic acid staining of monkey mesenchymal stem cells (MSCs)

The monkey MSCs expressed both $\alpha 2-3$ linked and $\alpha 2-6$ linked sialic acids on the surface. The filled histograms indicate cells as a control without addition of lectins. The lines indicate cells stained with specific lectins, and the broken lines indicate cells stained with specific lectins after treatment with sialidase to remove sialic acids on the surface of the cells. The cells treated with sialidase revealed a lower fluorescence intensity than did those without treatment with sialidase, which indicates that each lectin specifically recognize sialic acids on the cell surface.