

1 **Title:** The association between chronic deciduitis and preeclampsia.

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#### 41 **Highlights**

42 • Preeclampsia is frequently found in pregnant with chronic deciduitis.

43 • Chronic deciduitis is frequently found in pregnant with preeclamsia.

44 • Chronic deciduitis is related to poor perinatal outcomes.

45

46 **Key words:** chronic deciduitis, chronic endometritis, hypertensive disorders of

47 pregnancy, perinatal outcomes, preeclampsia

48

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50 DK, TH, AN, JK, TH; analyzed the data: AM, FK, SM, AN, JK, TH, RK; drafting the  
51 manuscript: AM, FK; substantively revised it: SM, ST, DK, TH, TA, RK, TM; final  
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53

54 **Ethics statement:**

55 The authors confirm that the ethical policies of the journal, as noted on the journal's  
56 author guidelines page. This research was approved by the Ethics Committee of Shiga  
57 University of Medical Science (R2020-074). The information on conducting the study  
58 was made public, and the opportunity for refusal was guaranteed as much as possible by  
59 optout as the Ethics Committee suggested.

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**71 Abstract**

72 Chronic deciduitis (CD) is slight inflammation of the decidua found during pregnancy.  
73 The cause of preeclampsia is thought to be placental hypoplasia, and various theories  
74 have been proposed to explain the detailed mechanism; however, its association with  
75 decidual inflammation is unclear. A retrospective case control study was conducted in a  
76 single university. Subjects were cases who delivered by cesarean section between January  
77 1, 2013 and June 30, 2020 and whose placentas were pathological assessed. CD was  
78 diagnosed by CD138 immunostaining of placental decidua tissue, and the perinatal  
79 prognosis and incidences of hypertensive disorder of pregnancy (HDP) and preeclmipsia  
80 were examined according to the presence or absence of CD. A logistic regression analysis  
81 was performed to evaluate the association between preeclampsia and 11 explanatory  
82 variables (10 patient or perinatal background factors and CD). The study population  
83 included 76 patients (non-CD, n=54; CD, n=22). The rate of preeclampsia was  
84 significantly higher in the CD group ( $P=0.0006$ ). Patients with CD gave birth at a  
85 significantly earlier gestational age ( $P=0.040$ ) with a lower birth weight ( $P=0.001$ ), and  
86 a higher rate of LFD ( $P=0.005$ ). The Apgar scores at 1 and 5 minutes and umbilical artery  
87 pH were lower ( $P=0.0003$ , 0.021 and 0.002, respectively) in the CD group. The logistic  
88 regression analysis revealed that CD was positively associated with preeclampsia. A  
89 retrospective examination of the placenta found that patients with CD had a significantly  
90 higher incidence of preeclampsia and CD is considered to be a factor that is associated  
91 with poor perinatal outcomes.

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95 **Main text**

96 **1. Introduction**

97 Chronic deciduitis (CD) is a type of long-term slight inflammation of the decidua that  
98 occurs during pregnancy (Khong et al., 2000) (Kim et al., 2015). Chronic microbial  
99 infection or immune mechanisms have been implicated in the etiology of CD (Kim et al.,  
100 2015), although the details of its pathophysiology remain unknown. CD is histologically  
101 diagnosed based on the presence of lymphocytes and plasma cells in the decidua(Khong  
102 et al., 2000) (Kimura et al., 2019) (Maroun et al., 2014). In particular, it has been reported  
103 that infiltration of plasma cells into the decidua results in a high severity of CD and is  
104 highly diagnostic for CD (Khong et al., 2000). CD is infrequently noticed by clinicians  
105 because it is histologically diagnosed using the removed placenta, but it can affect  
106 immunity and influence the establishment and maintenance of pregnancy. In fact, recent  
107 reports have revealed that CD is associated with miscarriage (Kaku et al., 2020), fetal  
108 growth restriction (Bendon and Miller, 1990), fetal death (Bendon and Miller, 1990), and  
109 preterm labor (Edmondson et al., 2009).

110 Hypertensive disorders of pregnancy (HDP) is a disease that causes hypertension,  
111 proteinuria and organ damage in pregnant women and results in preterm birth and fetal  
112 growth restriction, resulting in a poor perinatal prognosis for both the pregnant woman  
113 and newborn. Abnormal placentation is considered a major cause of HDP, especially  
114 preeclampsia. However, the detailed mechanism underlying abnormal placentation has  
115 not been clarified, although many theories, including oxidative stress, immunity, infection,  
116 genetic and environmental factors (Brown et al., 2018) (Burton et al., 2019) (Geldenhuys  
117 et al., 2018) (Nakashima et al., 2020) (Rana et al., 2019) (Saito, 2010) (Tsuda et al., 2019)  
118 (Wang et al., 2009), have been proposed.

119 We hypothesize that a long-lasting inflammation of the decidua of CD is associated with  
120 HDP and preeclampsia. The present study examined the relationship of CD with HDP and  
121 preeclampsia and evaluated the effect of CD on the perinatal outcomes.

122

## 123 **2. Methods**

### 124 **2.1. Ethics**

125 This study was approved by the Ethics Committee of Shiga University of Medical Science  
126 (R2020-074). Information about the study was made public, and the opportunity for  
127 refusal was guaranteed as much as possible by an optout method, as suggested by the  
128 Ethics Committee.

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### 130 **2.2. Subject**

131 The subjects were cases who delivered by cesarean section due to non-reassuring fetal  
132 status, fetal growth restriction, preterm delivery, fetal anomalies, and/or HDP between  
133 January 1, 2013 and June 30, 2020 and whose placentas were pathologically assessed.  
134 Cases at <22 weeks gestation, with premature rupture of the membranes, clinical  
135 chorioamnionitis, abruptio placenta and cases with abnormal morphological findings,  
136 such as placental tumor, were excluded from the study.

137

### 138 **2.3. Immunohistochemistry**

139 To diagnose CD, immunostaining of CD138 was performed to identify plasma cells of  
140 the decidua, as previously reported with slight modification ([Kaku et al., 2020](#)). In each  
141 case, the paraffin-embedded placental specimens containing the largest amount of  
142 decidua were used for the study. The specimens were cut into 4- $\mu$ m-thick sections, placed

143 on a 42°C water bath, and mounted on 3-aminopropyltriethoxysilane-coated slides (FRC-  
144 05; Matsunami, Osaka, Japan). Immunohistochemical staining reactions were performed  
145 using the DISCOVERY XT IHC Research Platform (F. Hoffmann-La Roche, Ltd.,  
146 Grenzacherstrasse, CH). Briefly, after deparaffinization and rehydration, the slides were  
147 placed in Cell Conditioning 1 (F. Hoffmann-La Roche, Ltd., Grenzacherstrasse, CH) and  
148 boiled at 98°C for 60 minutes. The slides were incubated with the monoclonal primary  
149 antibodies for CD138 (B-A38; Nichirei Corp., Tokyo, Japan) for 32 minutes at room  
150 temperature and incubated with the secondary antibody (Universal Secondary Antibody;  
151 F. Hoffmann-La Roche, Ltd., Grenzacherstrasse, CH) for 12 minutes at room temperature,  
152 followed by the development of DAB using the labeled streptavidin biotinylated antibody  
153 reaction.

154

#### 155 **2.4. Analyses**

156 The diagnosis of CD was performed by a blinded investigator. When one or more plasma  
157 cells stained with CD138 were found in the decidua of 10 random high-power fields  
158 (HPF), the patient was diagnosed with CD, as previously reported ([Kaku et al., 2020](#))  
159 ([Khong et al., 2000](#)). When findings of thrombosis, segmental avascular villi, and villous  
160 stromal-vascular karyorrhexis were detected according to the Amsterdam Placental  
161 workshop Group Consensus Statement ([Khong et al., 2016](#)), the diagnosis of fetal  
162 vascular malperfusion (FVM) was made. After blinding, one pathologist assessed the  
163 presence or absence of FVM, and then others analyzed the results.

164 The patients' characteristics, including age, gravidity, parity, body mass index (BMI),  
165 achievement of pregnancy by *in vitro* fertilization (IVF pregnancy), gestational age at  
166 delivery, birth weight, light for date infant status (LFD), Apgar scores at 1 and 5 minutes,

167 umbilical artery pH, presence of gestational diabetes mellitus (GDM) and HDP were  
168 extracted from the patients' records. When the HDP was present, the presence of severe  
169 HDP and type of HDP (preeclampsia, gestational hypertension and chronic hypertension)  
170 were extracted. When preeclampsia existed, the presence of early onset-type  
171 preeclampsia was ascertained.

172 Then, the patients were divided into two groups according to the presence (CD group) or  
173 absence (non-CD group) of CD. Each item was compared between the groups.

174 In addition, patients diagnosed with preeclampsia were divided into CD and non-CD  
175 groups and the parameters, including FVM, were compared between the groups.

176 Next, the patients were classified into those who developed preeclampsia (preeclampsia  
177 group) and those who did not develop any type of HDP (non-HDP group). Each parameter  
178 was compared between the preeclampsia and non-HDP groups.

179 We calculated the number of patients required for enrollment. We referred a report which  
180 showed that 32.1% of pregnant women with chronic endometritis (CE) developed  
181 preeclampsia when CE was not treated before pregnancy (Taranovska Ocapital O et al.,  
182 2020). HDP generally occurred at a rate of 5–10% (Hutcheon et al., 2011) (Li et al., 2021).  
183 However, this general population included patients with CE. CE has been reported to be  
184 observed in the uterine endometrium of 50–70% of women with a history of recurrent  
185 pregnancy loss (McQueen et al., 2015) (Zolghadri et al., 2011). Based on these references,  
186 we set the proportion of HDP unrelated to CE to 3.5%. Thus, we selected 0.05 for  $\alpha$ , 0.8  
187 for power  $\beta$ , 0.035 for  $P_0$  (the probability for a control patient), and 0.32 for  $P_1$  (the  
188 probability in an experimental subject). The calculation revealed that 21 experimental  
189 subjects and 53 control subjects were required. Alternatively, we referred to another report  
190 (Gundogan et al., 2010), which showed that the incidence rates of CD and HDP were



191 higher in an egg donation group in comparison to controls. CD was found in 42.4%  
192 (14/33) of the placentas in the egg donation group and 1.7% (2/60) of the placentas in the  
193 control group. Thus, we selected 0.017 for P0, and 0.424 for P1. The calculation resulted  
194 in sample sizes of 20 cases for the control group and 15 cases for the affected group.

195 The statistical analyses were performed using Graph Pad Prism 6 (GraphPad Software  
196 Inc., La Jolla, CA) and SPSS version 25 (SPSS Inc, Chicago, IL). Each dataset was  
197 analyzed for normal distribution using D'Agostino's *K*-squared test, and Student's *t*-test  
198 or the non-parametric Mann-Whitney *U*-test was used depending on the distribution  
199 pattern.

200 When the distribution pattern was judged as parametric, the result was shown as the  
201 mean  $\pm$  SD. When the data were judged to show a non-parametric distribution, the results  
202 were shown as the median (interquartile range).

203 The statistical comparison of IVF pregnancy, LFD and HDP in the CD and non-CD  
204 group and IVF pregnancy, CD and LFD in the preeclampsia and non-HDP group was  
205 performed using Fisher's exact test.

206 A stepwise logistic regression analysis of factors associated with preeclampsia or CD  
207 was performed to examine the independent effect of each of the following variables: age  
208  $\geq 40$  years, BMI  $\geq 25$ , multigravida, multiparity, IVF pregnancy, multiple pregnancy,  
209 preterm birth, CD, preeclampsia, LFD, Apgar score at 5 minutes  $\leq 6$ , umbilical artery pH  
210  $< 7.1$ , and sex of the child. *P* values of  $< 0.05$  were considered to indicate statistical  
211 significance.

212

### 213 **3. Results**

#### 214 **3.1. Immunohistochemistry**

215 Plasma cells infiltrating the decidua were identified by immunostaining for CD138  
216 (Figure 1). Chorionic cells were stained with CD138 in all cases. When plasma cells were  
217 detected in the decidua, they often formed clusters.

218

### 219 **3.2. Clinical parameters in the non-CD and CD groups**

220 There were 54 patients in the non-CD group and 22 in the CD group. The patients'  
221 background clinical characteristics are shown in Table 1. There were no differences in  
222 age, BMI, gravidity, parity, or IVF pregnancy between the non-CD and CD groups. The  
223 rate of HDP was significantly higher in the CD group ( $P=0.002$ ). Among the subtypes of  
224 HDP, only preeclampsia was found to have a significantly higher incidence in CD  
225 ( $P=0.0006$ ). Gestational hypertension was found only in non-CD, although the difference  
226 in the incidence was not significant. Chronic hypertension was not found in either  
227 condition. The correlation between preeclampsia and CD was stronger than that between  
228 HDP and CD. The rate of severe HDP was significantly higher in the CD group  
229 ( $P=0.0008$ ). There was no difference in the rate of GDM. Patients with CD gave birth at  
230 a significantly earlier gestational age ( $P=0.040$ ), with a smaller birth weight ( $P=0.001$ ),  
231 and with a higher rate of LFD ( $P=0.005$ ). Furthermore, in the CD group, the Apgar score  
232 at 1 minute, Apgar score at 5 minutes, and umbilical cord arterial blood pH were lower  
233 ( $P=0.0003$ ,  $0.021$  and  $0.002$ ).

234 Since gestational hypertension was only found in the non-CD group, chronic  
235 hypertension was not found in both groups and the correlation between preeclampsia and  
236 CD was stronger than that between HDP and CD, subsequent studies were limited to  
237 preeclampsia, with the preeclampsia group compared to the non-HDP group

238

### 239 3.3. Results of a stepwise logistic regression analysis with preeclampsia

240 A logistic regression analysis was performed with preeclampsia as the objective variable  
241 (Table 2). The explanatory variables were age  $\geq 40$  years, BMI  $\geq 25$ , multigravida,  
242 multiparity, IVF pregnancy, multiple pregnancy, preterm birth, CD, LFD, Apgar score at  
243 5 minutes  $\leq 6$ , umbilical artery pH  $< 7.1$ , and sex of the child. The following variables were  
244 found to be related to preeclampsia: age  $\geq 40$  years, BMI  $\geq 25$ , multigravida, and CD. The  
245 results of the Hosmer and Lemeshow tests were  $P=0.957$  and the discriminative  
246 predictive value was 81.7%.

247

### 248 3.4. Clinical parameters of the non-CD and CD groups with preeclampsia

249 In addition, a sub-analysis was performed to show the differences between patients with  
250 and without CD in the preeclampsia group (Table 3). In this sub-analysis, there were no  
251 differences in age, gravidity, parity, BMI, or IVF pregnancy between the patients with  
252 and without CD. There was no significant differences between the two groups in the  
253 gestational age at delivery, the rate of preterm birth, the rate of LFD, or Apgar score at 5  
254 minutes.

255 In this sub-analysis, the patients with CD were found to have a significantly lower birth  
256 weight ( $P=0.042$ ). Furthermore, in this sub-analysis, the Apgar score at 1 minute tended  
257 to be lower, and the umbilical artery pH was significantly lower in patients with CD  
258 ( $P=0.034$ ). CD was more frequently found in patients with early-onset type preeclampsia.  
259 The rate of FVM was not different.

260

### 261 3.5. Clinical parameters of the non- HDP and preeclampsia groups

262 The patients were classified into the non- HDP group ( $n=41$ ) and the preeclampsia group

263 (n=31). The patients' background clinical characteristics are shown in Table 4. The  
264 preeclampsia group had significantly higher age and BMI values ( $P=0.047$  and  $P=0.013$ ,  
265 respectively) in comparison to the non- HDP group. Gravidity was significantly lower  
266 ( $P=0.031$ ) in the preeclampsia group than in the non- HDP group and parity tended to be  
267 lower ( $P=0.073$ ) in the preeclampsia group. The rate of IVF pregnancy tended to be  
268 higher in the preeclampsia group. There was no difference in the gestational age at  
269 delivery; however, the birth weight was lower ( $P=0.003$ ) and the rate of LFD was higher  
270 ( $P=0.036$ ) in the preeclampsia group. There were no differences in the Apgar scores at 1  
271 and 5 minutes; however, the umbilical artery pH was lower ( $P=0.038$ ) in the preeclampsia  
272 group.

273 CD was detected in 51.6% of the patients in the preeclampsia group and 17.1% of the  
274 patients in the non- HDP group ( $P=0.0024$ ) (Table 4). The comparison of the number of  
275 CD138-positive cells in 10 random HPFs between the preeclampsia and non- HDP groups  
276 revealed that the number of CD138-positive cells was significantly higher in the  
277 preeclampsia group ( $P=0.0005$ ) (Figure 2).

278

### 279 **3.6. Results of a stepwise logistic regression analysis with CD**

280 In the logistic regression analysis with CD as the objective variable (Table 2), the  
281 explanatory variables were age  $\geq 40$  years, BMI  $\geq 25$ , multigravida, multiparity, IVF  
282 pregnancy, multiple pregnancy, preterm birth, preeclampsia, LFD, Apgar score at 5  
283 minutes  $\leq 6$ , umbilical artery pH  $< 7.1$ , and sex of the child. The variables multiparity,  
284 preeclampsia, and LFD were found to be related to CD. The results of the Hosmer and  
285 Lemeshow tests were  $P=0.807$ , and the discriminative predictive value was 78.9%.

286

#### 287 4. Discussion

288 In the present study, the incidence of HDP in the CD group was found to be higher than  
289 that in the non-CD group; preeclampsia was the only type of HDP for which the incidence  
290 was significantly higher in the CD group. Furthermore, the incidence of CD in the  
291 preeclampsia group was higher than that in the non-HDP group. To the best of our  
292 knowledge, this is the first report to show an association between the presence of CD and  
293 the development of preeclampsia.

294 On the other hand, the present study also found that CD is a factor that worsens  
295 perinatal outcomes. In the past, CD was reported to be associated with fetal growth  
296 restriction (Bendon and Miller, 1990), intrauterine fetal death (Bendon and Miller, 1990)  
297 and preterm birth (Edmondson et al., 2009). Our data support these previous reports in  
298 terms of the higher rates of LFD and preterm birth.

299

300 CD is characterized by slight inflammation of the placental decidua. Regarding the  
301 relationship between inflammation and HDP, lipopolysaccharide has been reported to  
302 induce cytokine production and reduce EVT invasion (Anton et al., 2012). Moreover,  
303 numerous cytokines secreted by both trophoblast cells and decidua play a role in  
304 promoting and suppressing the infiltration of trophoblast cells during implantation  
305 (Sharma et al., 2016). We speculated that inflammation of the decidua might affect  
306 trophoblast invasion and spiral artery remodeling via the modulation of the immunity and  
307 the endometrial/decidual cell function (Di Pietro et al., 2013) (Kitazawa et al., 2021)  
308 (Matteo et al., 2009) (Pollheimer et al., 2018) (Tang et al., 2015) (Wu et al., 2017).  
309 Recently, we reported that patients in whom CD is diagnosed based on the examination  
310 a miscarriage specimen have a high rate of chronic endometritis (CE) (Kaku et al., 2020),

311 which involves slight inflammation of the endometrium (Kimura et al., 2019) (Kitaya et  
312 al., 2016). Thus, a certain rate of CE before pregnancy peppepersists after pregnancy  
313 and becomes CD.

314 Moreover, patients in whom CE is detected before pregnancy have been reported to have  
315 a higher incidence of miscarriage (Morimune et al., 2021) , preterm birth (Morimune et  
316 al., 2021) and preeclampsia (Taranovska Ocapital O et al., 2020). Taken together, we  
317 speculated that endometrial inflammation may persist in the early-stage decidua in certain  
318 cases and that persistent inflammation might inhibit spiral artery remodeling, resulting in  
319 the development of preeclampsia.

320 On the other hand, it is possible that abnormal immune tolerance between the mother and  
321 infant caused inflammation and CD. The egg donor IVF placenta, in which the mother  
322 and embryo are completely immunogenetically unrelated, has been reported to have a  
323 significantly higher incidence of CD than the non-donor IVF placenta (Gundogan et al.,  
324 2010). These differences were suggested to result from differentially modulated immune  
325 activity at the maternal-fetal interface of egg donor pregnancies (Gundogan et al., 2010,  
326 Saito et al., 2016). Preterm births are reported to be more common in egg donation  
327 pregnancies, suggesting that CD may be associated with preterm birth (Gundogan et al.,  
328 2010). Previous studies have demonstrated that egg donor pregnancies are associated with  
329 an increased risk of preeclampsia (Blazquez et al., 2016) (Masoudian et al., 2016).  
330 Abnormal immune tolerance between mothers and infants is strongly supported as a cause  
331 of preeclampsia. We speculated that a host versus graft rejection phenomenon or an effort  
332 to suppress rejection causes CD, and that these immunological abnormalities also cause  
333 preeclampsia (Morita et al., 2020) (Redman and Sargent, 2010) (Tsuda et al., 2018).

334 It is now clear that bacterial flora is present in many organs of adult humans. The

335 predominant idea at present is that the bacterial flora is present in the placenta, although  
336 some reports have denied such a thing. However, the presence of bacterial flora in the  
337 endometrium, regardless of the presence of bacteria in the placenta, has been confirmed  
338 not only via next-generation sequencing (Chen et al., 2017) but also in simple culture  
339 studies (Benner et al., 2018) (Cowling et al., 1992) (Moller et al., 1995). Given these  
340 results, we believe that plasma cells in the decidua could develop due to antigens in the  
341 maternal endometrium or its decidualized endometrium although it is possible that they  
342 develop due to certain placental antigens which derived from the fetus or antigens existing  
343 in the placenta such as bacteria.

344

345 The strength of this study is that it is the first report to clearly reveal the association  
346 between preeclampsia and CD. However, the present study is associated with some  
347 limitations. This was a retrospective study and subjects were only selected when the  
348 placenta was assessed, after the extraction of cases involving delivery by cesarean section  
349 due to non-reassuring fetal status, fetal growth restriction, preterm delivery, fetal anomaly,  
350 and/or HDP. Thus, although we confirmed the results by a logistic regression analysis,  
351 there was a selection bias.

352

353 In the future, it is expected that preeclampsia might be fundamentally treated by  
354 elucidating the pathophysiology of CD. In particular, in a certain number of cases, CD  
355 was derived from CE. In cases involving CE, the live birth rate has been reported to be  
356 increased by antibiotic treatment before pregnancy (Cicinelli et al., 2015) (Kitaya et al.,  
357 2017). In the future study, it would be worthwhile evaluating whether the curative  
358 treatment of CE before pregnancy can prevent the onset of preeclampsia.

359 In conclusion, based on the retrospective examination of placenta, we found that patients  
360 with CD had a significantly higher incidence of preeclampsia and CD is considered to be  
361 a factor that is associated with poor perinatal outcomes.

362

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366

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368 None of the authors has any conflict of interest related to this manuscript.

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382 **References**

- 383 ANTON, L., BROWN, A. G., PARRY, S. & ELOVITZ, M. A. 2012. Lipopolysaccharide  
384 induces cytokine production and decreases extravillous trophoblast invasion  
385 through a mitogen-activated protein kinase-mediated pathway: possible  
386 mechanisms of first trimester placental dysfunction. *Hum Reprod*, 27, 61-72.
- 387 BENDON, R. W. & MILLER, M. 1990. Routine pathological examination of placentae  
388 from abnormal pregnancies. *Placenta*, 11, 369-70.
- 389 BENNER, M., FERWERDA, G., JOOSTEN, I. & VAN DER MOLEN, R. G. 2018. How  
390 uterine microbiota might be responsible for a receptive, fertile endometrium. *Hum*  
391 *Reprod Update*, 24, 393-415.
- 392 BLAZQUEZ, A., GARCIA, D., RODRIGUEZ, A., VASSENA, R., FIGUERAS, F. &  
393 VERNAEVE, V. 2016. Is oocyte donation a risk factor for preeclampsia? A  
394 systematic review and meta-analysis. *J Assist Reprod Genet*, 33, 855-63.
- 395 BROWN, M. A., MAGEE, L. A., KENNY, L. C., KARUMANCHI, S. A., MCCARTHY,  
396 F. P., SAITO, S., HALL, D. R., WARREN, C. E., ADOYI, G., ISHAKU, S. &  
397 INTERNATIONAL SOCIETY FOR THE STUDY OF HYPERTENSION IN, P.  
398 2018. Hypertensive Disorders of Pregnancy: ISSHP Classification, Diagnosis, and  
399 Management Recommendations for International Practice. *Hypertension*, 72, 24-  
400 43.
- 401 BURTON, G. J., REDMAN, C. W., ROBERTS, J. M. & MOFFETT, A. 2019. Pre-  
402 eclampsia: pathophysiology and clinical implications. *BMJ*, 366, l2381.
- 403 CHEN, C., SONG, X., WEI, W., ZHONG, H., DAI, J., LAN, Z., LI, F., YU, X., FENG,  
404 Q., WANG, Z., XIE, H., CHEN, X., ZENG, C., WEN, B., ZENG, L., DU, H.,  
405 TANG, H., XU, C., XIA, Y., XIA, H., YANG, H., WANG, J., WANG, J.,  
406 MADSEN, L., BRIX, S., KRISTIANSEN, K., XU, X., LI, J., WU, R. & JIA, H.  
407 2017. The microbiota continuum along the female reproductive tract and its  
408 relation to uterine-related diseases. *Nat Commun*, 8, 875.
- 409 CICINELLI, E., MATTEO, M., TINELLI, R., LEPERA, A., ALFONSO, R.,  
410 INDRACCOLO, U., MARROCHELLA, S., GRECO, P. & RESTA, L. 2015.  
411 Prevalence of chronic endometritis in repeated unexplained implantation failure  
412 and the IVF success rate after antibiotic therapy. *Hum Reprod*, 30, 323-30.
- 413 COWLING, P., MCCOY, D. R., MARSHALL, R. J., PADFIELD, C. J. & REEVES, D.  
414 S. 1992. Bacterial colonization of the non-pregnant uterus: a study of pre-  
415 menopausal abdominal hysterectomy specimens. *Eur J Clin Microbiol Infect Dis*,  
416 11, 204-5.
- 417 DI PIETRO, C., CICINELLI, E., GUGLIELMINO, M. R., RAGUSA, M., FARINA, M.,

- 418 PALUMBO, M. A. & CIANCI, A. 2013. Altered transcriptional regulation of  
419 cytokines, growth factors, and apoptotic proteins in the endometrium of infertile  
420 women with chronic endometritis. *Am J Reprod Immunol*, 69, 509-17.
- 421 EDMONDSON, N., BOCKING, A., MACHIN, G., RIZEK, R., WATSON, C. &  
422 KEATING, S. 2009. The prevalence of chronic deciduitis in cases of preterm labor  
423 without clinical chorioamnionitis. *Pediatr Dev Pathol*, 12, 16-21.
- 424 GELDENHUYS, J., ROSSOUW, T. M., LOMBAARD, H. A., EHLERS, M. M. & KOCK,  
425 M. M. 2018. Disruption in the Regulation of Immune Responses in the Placental  
426 Subtype of Preeclampsia. *Front Immunol*, 9, 1659.
- 427 GUNDOGAN, F., BIANCHI, D. W., SCHERJON, S. A. & ROBERTS, D. J. 2010.  
428 Placental pathology in egg donor pregnancies. *Fertil Steril*, 93, 397-404.
- 429 HUTCHEON, J. A., LISONKOVA, S. & JOSEPH, K. S. 2011. Epidemiology of pre-  
430 eclampsia and the other hypertensive disorders of pregnancy. *Best Pract Res Clin*  
431 *Obstet Gynaecol*, 25, 391-403.
- 432 KAKU, S., KUBO, T., KIMURA, F., NAKAMURA, A., KITAZAWA, J., MORIMUNE,  
433 A., TAKAHASHI, A., TAKEBAYASHI, A., TAKASHIMA, A., KUSHIMA, R. &  
434 MURAKAMI, T. 2020. Relationship of chronic endometritis with chronic  
435 deciduitis in cases of miscarriage. *BMC Womens Health*, 20, 114.
- 436 KHONG, T. Y., BENDON, R. W., QURESHI, F., REDLINE, R. W., GOULD, S.,  
437 STALLMACH, T., LIPSETT, J. & STAPLES, A. 2000. Chronic deciduitis in the  
438 placental basal plate: definition and interobserver reliability. *Hum Pathol*, 31, 292-  
439 5.
- 440 KHONG, T. Y., MOONEY, E. E., ARIEL, I., BALMUS, N. C., BOYD, T. K.,  
441 BRUNDLER, M. A., DERRICOTT, H., EVANS, M. J., FAYE-PETERSEN, O.  
442 M., GILLAN, J. E., HEAZELL, A. E., HELLER, D. S., JACQUES, S. M.,  
443 KEATING, S., KELEHAN, P., MAES, A., MCKAY, E. M., MORGAN, T. K.,  
444 NIKKELS, P. G., PARKS, W. T., REDLINE, R. W., SCHEIMBERG, I.,  
445 SCHOOTS, M. H., SEBIRE, N. J., TIMMER, A., TUROWSKI, G., VAN DER  
446 VOORN, J. P., VAN LIJNSCHOTEN, I. & GORDIJN, S. J. 2016. Sampling and  
447 Definitions of Placental Lesions: Amsterdam Placental Workshop Group  
448 Consensus Statement. *Arch Pathol Lab Med*, 140, 698-713.
- 449 KIM, C. J., ROMERO, R., CHAEMSAITHONG, P. & KIM, J. S. 2015. Chronic  
450 inflammation of the placenta: definition, classification, pathogenesis, and clinical  
451 significance. *Am J Obstet Gynecol*, 213, S53-69.
- 452 KIMURA, F., TAKEBAYASHI, A., ISHIDA, M., NAKAMURA, A., KITAZAWA, J.,  
453 MORIMUNE, A., HIRATA, K., TAKAHASHI, A., TSUJI, S., TAKASHIMA, A.,

- 454 AMANO, T., TSUJI, S., ONO, T., KAKU, S., KASAHARA, K., MORITANI, S.,  
455 KUSHIMA, R. & MURAKAMI, T. 2019. Review: Chronic endometritis and its  
456 effect on reproduction. *J Obstet Gynaecol Res*, 45, 951-960.
- 457 KITAYA, K., MATSUBAYASHI, H., TAKAYA, Y., NISHIYAMA, R., YAMAGUCHI,  
458 K., TAKEUCHI, T. & ISHIKAWA, T. 2017. Live birth rate following oral  
459 antibiotic treatment for chronic endometritis in infertile women with repeated  
460 implantation failure. *Am J Reprod Immunol*, 78.
- 461 KITAYA, K., MATSUBAYASHI, H., YAMAGUCHI, K., NISHIYAMA, R., TAKAYA,  
462 Y., ISHIKAWA, T., YASUO, T. & YAMADA, H. 2016. Chronic Endometritis:  
463 Potential Cause of Infertility and Obstetric and Neonatal Complications. *Am J*  
464 *Reprod Immunol*, 75, 13-22.
- 465 KITAZAWA, J., KIMURA, F., NAKAMURA, A., MORIMUNE, A., HANADA, T.,  
466 AMANO, T., TSUJI, S., KASAHARA, K., SATOOKA, H., HIRATA, T.,  
467 KUSHIMA, R. & MURAKAMI, T. 2021. Alteration in endometrial helper T-cell  
468 subgroups in chronic endometritis. *Am J Reprod Immunol*, 85, e13372.
- 469 LI, X., ZHANG, W., LIN, J., LIU, H., YANG, Z., TENG, Y., HUANG, J., PENG, Q., LIN,  
470 X., ZHANG, J., XIE, L., XIE, Y., LI, Y., LUO, J., DUAN, W., CHEN, J. & DUAN,  
471 S. 2021. Hypertensive disorders of pregnancy and risks of adverse pregnancy  
472 outcomes: a retrospective cohort study of 2368 patients. *J Hum Hypertens*, 35, 65-  
473 73.
- 474 LUCAS, D N., YENTIS, S M., KINSELLA, S M., HOLDCROFT, A., MAY, A E., WEE,  
475 M., ROBINSON, P N. 2000. Urgency of caesarean section: a new classification. *J*  
476 *R Soc Med*, 93, 346-50.
- 477 MAROUN, L. L., MATHIESEN, L., HEDEGAARD, M., KNUDSEN, L. E. & LARSEN,  
478 L. G. 2014. Pathologic evaluation of normal and perfused term placental tissue.  
479 *Pediatr Dev Pathol*, 17, 330-8.
- 480 MASOUDIAN, P., NASR, A., DE NANASSY, J., FUNG-KEE-FUNG, K.,  
481 BAINBRIDGE, S. A. & EL DEMELLAWY, D. 2016. Oocyte donation  
482 pregnancies and the risk of preeclampsia or gestational hypertension: a systematic  
483 review and metaanalysis. *Am J Obstet Gynecol*, 214, 328-39.
- 484 MATTEO, M., CICINELLI, E., GRECO, P., MASSENZIO, F., BALDINI, D.,  
485 FALAGARIO, T., ROSENBERG, P., CASTELLANA, L., SPECCHIA, G. &  
486 LISO, A. 2009. Abnormal pattern of lymphocyte subpopulations in the  
487 endometrium of infertile women with chronic endometritis. *Am J Reprod Immunol*,  
488 61, 322-9.
- 489 MCQUEEN, D. B., PERFETTO, C. O., HAZARD, F. K. & LATHI, R. B. 2015.

- 490 Pregnancy outcomes in women with chronic endometritis and recurrent  
491 pregnancy loss. *Fertil Steril*, 104, 927-931.
- 492 MOLLER, B. R., KRISTIANSEN, F. V., THORSEN, P., FROST, L. & MOGENSEN, S.  
493 C. 1995. Sterility of the uterine cavity. *Acta Obstet Gynecol Scand*, 74, 216-9.
- 494 MORIMUNE, A., KIMURA, F., NAKAMURA, A., KITAZAWA, J., TAKASHIMA, A.,  
495 AMANO, T., KAKU, S., MORITANI, S., KUSHIMA, R. & MURAKAMI, T.  
496 2021. The effects of chronic endometritis on the pregnancy outcomes. *Am J*  
497 *Reprod Immunol*, 85, e13357.
- 498 MORITA, K., TSUDA, S., KOBAYASHI, E., HAMANA, H., TSUDA, K., SHIMA, T.,  
499 NAKASHIMA, A., USHIJIMA, A., KISHI, H. & SAITO, S. 2020. Analysis of  
500 TCR Repertoire and PD-1 Expression in Decidual and Peripheral CD8(+) T Cells  
501 Reveals Distinct Immune Mechanisms in Miscarriage and Preeclampsia. *Front*  
502 *Immunol*, 11, 1082.
- 503 NAKASHIMA, A., SHIMA, T., AOKI, A., KAWAGUCHI, M., YASUDA, I., TSUDA,  
504 S., YONEDA, S., YAMAKI-USHIJIMA, A., CHENG, S., SHARMA, S. &  
505 SAITO, S. 2020. Placental autophagy failure: A risk factor for preeclampsia. *J*  
506 *Obstet Gynaecol Res*.
- 507 POLLHEIMER, J., VONDRA, S., BALTAYEVA, J., BERISTAIN, A. G. & KNOFLER,  
508 M. 2018. Regulation of Placental Extravillous Trophoblasts by the Maternal  
509 Uterine Environment. *Front Immunol*, 9, 2597.
- 510 RANA, S., LEMOINE, E., GRANGER, J. P. & KARUMANCHI, S. A. 2019.  
511 Preeclampsia: Pathophysiology, Challenges, and Perspectives. *Circ Res*, 124,  
512 1094-1112.
- 513 REDMAN, C. W. & SARGENT, I. L. 2010. Immunology of pre-eclampsia. *Am J Reprod*  
514 *Immunol*, 63, 534-43.
- 515 SAITO, S. 2010. Th17 cells and regulatory T cells: new light on pathophysiology of  
516 preeclampsia. *Immunol Cell Biol*, 88, 615-7.
- 517 SAITO, S., NAKABAYASHI, Y., NAKASHIMA, A., SHIMA, T. & YOSHINO, O. 2016.  
518 A new era in reproductive medicine: consequences of third-party oocyte donation  
519 for maternal and fetal health. *Semin Immunopathol*, 38, 687-697.
- 520 SHARMA, S., GODBOLE, G. & MODI, D. 2016. Decidual Control of Trophoblast  
521 Invasion. *Am J Reprod Immunol*, 75, 341-50.
- 522 TANG, M. X., HU, X. H., LIU, Z. Z., KWAK-KIM, J. & LIAO, A. H. 2015. What are the  
523 roles of macrophages and monocytes in human pregnancy? *J Reprod Immunol*,  
524 112, 73-80.
- 525 TARANOVSKA OCAPITAL O, C., LIKHACHOV VCAPITAL KA, C.,

- 526 DOBROVOLSKA L, CAPITAL EM, C., MAKAROV, O. G. & SHYMANSKA, Y.  
527 V. 2020. The Role of Secreting Function of Decidua in the Development of  
528 Complications of Gestation Process in Pregnant Women with a Past History of  
529 Chronic Endometritis. *Wiad Lek*, 73, 2416-2420.
- 530 TSUDA, S., NAKASHIMA, A., SHIMA, T. & SAITO, S. 2019. New Paradigm in the  
531 Role of Regulatory T Cells During Pregnancy. *Front Immunol*, 10, 573.
- 532 TSUDA, S., ZHANG, X., HAMANA, H., SHIMA, T., USHIJIMA, A., TSUDA, K.,  
533 MURAGUCHI, A., KISHI, H. & SAITO, S. 2018. Clonally Expanded Decidual  
534 Effector Regulatory T Cells Increase in Late Gestation of Normal Pregnancy, but  
535 Not in Preeclampsia, in Humans. *Front Immunol*, 9, 1934.
- 536 WANG, A., RANA, S. & KARUMANCHI, S. A. 2009. Preeclampsia: the role of  
537 angiogenic factors in its pathogenesis. *Physiology (Bethesda)*, 24, 147-58.
- 538 WU, D., KIMURA, F., ZHENG, L., ISHIDA, M., NIWA, Y., HIRATA, K.,  
539 TAKEBAYASHI, A., TAKASHIMA, A., TAKAHASHI, K., KUSHIMA, R.,  
540 ZHANG, G., MURAKAMI, T. 2017. Chronic endometritis modifies  
541 decidualization in human endometrial stromal cells. *Reproductive Biology and  
542 Endocrinology*, 15.
- 543 ZOLGHADRI, J., MOMTAHAN, M., AMINIAN, K., GHAFFARPASAND, F. &  
544 TAVANA, Z. 2011. The value of hysteroscopy in diagnosis of chronic  
545 endometritis in patients with unexplained recurrent spontaneous abortion. *Eur J  
546 Obstet Gynecol Reprod Biol*, 155, 217-20.
- 547  
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562 **Figure Legends**

563 **Figure 1. Immunohistochemistry of CD138 for placenta.**

564 a. b. Plasma cells in the decidua immunostained by CD138 are detected in chronic  
565 deciduitis.

566 c. d. Plasma cells in the decidua are not detected in non-chronic deciduitis.

567 a. c. Bar = 500  $\mu$ m

568 b. d. Bar = 100  $\mu$ m

569

570 **Figure 2. The number of CD138-positive cells in 10 HPFs.**

571 The number of CD138-positive cells in 10 random HPFs was significantly higher in the  
572 preeclampsia group than in the non- HDP group.

573 There was a case with a markedly high number of CD138-positive cells. Five sections  
574 from different placental parts were pathologically examined to confirm the result  
575 (Supplemental figure 1) . As results, over 100 CD138-positive cells in 10 random HPFs  
576 were found to have infiltrated into the decidua in all specimens. This was a case of  
577 preeclampsia with delivery at 32 weeks' gestation in which the fetal heart rate pattern  
578 rapidly deteriorated. Among all cases examined in the present study, this was the only one  
579 for whom category-1 emergency caesarean section(Lucas et al., 2000) was performed in  
580 the preterm period. The infant was LFD, and the Apgar scores at 1 and 5 minutes (1 point,  
581 5 points) and umbilical arterial blood pH (6.882) were extremely low. This result might  
582 therefore indicate the serious effects of CD138-positive cells in the decidua on the  
583 pathophysiology of HDP and/or the prognosis of the affected fetus.

584

585 **Supplemental figure 1. Immunohistochemistry of CD138 for the placenta of the case**

586 **with markedly high numbers of CD138-positive cells.**

587 Over 100 CD138-positive cells in 10 random HPFs were found to have infiltrated into the

588 decidua of the placenta preserved in paraffin-embedded specimens.

589 a. b. c. d. e. Bar = 500  $\mu\text{m}$

590 f. Bar = 100  $\mu\text{m}$

1 **Table 1. Characteristics and delivery outcomes in the non-CD and CD groups**

	non-CD N=54	CD N=22	P value
Age (years)	33.98±5.11	33.82±5.73	NS
Gravity, median (IQR)	2.00 (1.00-2.00)	1.50 (1.00-2.00)	NS
Parity, median (IQR)	0 (0-1.00)	0 (0-1.00)	NS
BMI (kg/m <sup>2</sup> )	22.19±3.88	22.05±3.66	NS
IVF pregnancy	31.5% (17/54)	27.2% (6/22)	NS
HDP	35.2% (19/54)	72.7% (16/22)	<.01
Preeclampsia	27.8% (15/54)	72.7% (16/22)	<.001
Gestational hypertension	7.4% (4/54)	0% (0/22)	NS
Chronic hypertension	0% (0/54)	0% (0/22)	NS
Severe HDP	29.6% (16/54)	72.7% (16/22)	<.001
GDM	9.2% (5/54)	9.0% (2/22)	NS
Gestational age at delivery	35.30±2.57	33.73±3.67	<.05
Birth weight (g)	2270±645.28	1752±596.24	0.001
LFD	26.9% (18/67)	60.9% (14/23)	<.01
Apgar score at 1 minute	7.48 ±1.19	5.96 ±2.56	<.001
Apgar score at 5 minutes, median (IQR)	9.00 (9.00-9.00)	9.00 (8.00-9.00)	<.03
Umbilical artery pH, median (IQR)	7.313 (7.281-7.343)	7.281 (7.206- 7.301)	<.01

2 BMI, Body mass index; CD, Chronic deciduitis; GDM, Gestational diabetes mellitus;

3 HDP, Hypertensive disorders of pregnancy; IVF, *In vitro* fertilization; LFD, Light-for-

4 date infant; NS, Not significant

5



6 **Table 2. Logistic regression analysis of factors associated with preeclampsia and CD**

	Variable	Odds ratio	95% CI	P value
Preeclampsia	Age $\geq$ 40 years	57.3	3.7-877.7	<.01
	BMI $\geq$ 25	16.36	1.8-147.5	<.03
	Multigravida	0.08	0.01-0.4	<.01
	CD	12.1	1.9-73.7	<.01
CD	Multiparity	4.9	1.1-20.8	<.03
	Preeclampsia	7.7	1.9-30.2	<.01
	LFD	5.9	1.6-21.3	<.01

7 BMI, Body mass index; CD, Chronic deciduitis; CI, Confidence interval; LFD, Light-for-

8 date infant.

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12 **Table 3. Characteristics and outcomes of the preeclampsia patients in the non-CD**  
 13 **and CD groups .**

	non-CD N=15	CD N=16	P value
Age (y)	36.80±4.32	34.25±6.46	NS
Gravity	1.53±0.63	1.81±1.33	NS
Parity, median (IQR)	0.0 (0.0-0.0)	0.0 (0.0-1.0)	NS
BMI (kg/m <sup>2</sup> )	24.94±5.30	22.37±4.04	NS
IVF pregnancy	40.0% (6/15)	37.5% (6/16)	NS
Gestational age at delivery	35.07±3.32	33.13±4.06	NS
Preterm birth	53.3% (8/15)	81.2% (13/16)	NS
Birth weight (g)	2116±816.70	1577±562.23	<.05
LFD	40.0% (6/15)	62.5% (10/16)	NS
Apgar score at 1 minute	7.53±0.91	6.18±2.65	<.1
Apgar score at 5 minutes, median (IQR)	9.00 (9.00-9.00)	9.00 (8.25-9.00)	NS
Umbilical artery pH	7.306±0.04	7.228±0.13	<.05
Early-onset type PE	40.0% (6/15)	75.0% (12/16)	<.1
FVM	60.0% (9/15)	87.5% (14/16)	NS

14 BMI, Body mass index; CD, Chronic deciduitis; FVM, Fetal vascular malperfusion; IVF,

15 *In vitro* fertilization; LFD, Light-for-date infant; NS, Not significant.

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17 **Table 4. Characteristics and delivery outcomes of the non-HDP and preeclampsia**  
 18 **groups**

	non- HDP N=41	preeclampsia N=31	P value
Age (y)	32.8±4.91	35.4±5.65	<.05
Gravidity	2.34±1.37	1.67±1.07	<.05
Parity, median (IQR)	0.0 (0.0-1.0)	0.0 (0.0-1.0)	<.1
BMI (kg/m <sup>2</sup> )	21.34 ±2.56	23.61±4.85	<.03
IVF pregnancy	26.8% (11/41)	38.7% (12/31)	<.1
Gestational age at delivery	35.34±2.17	34.06±3.86	NS
Birth weight (g)	2280±575.56	1838±745.10	<.01
LFD	27.8% (15/54)	51.6% (16/31)	<.05
Apgar score at 1 minute, median (IQR)	8.00 (6.00-8.00)	8.00 (7.00-8.00)	NS
Apgar score at 5 minutes, median (IQR)	9.00 (8.00-9.00)	9.00 (8.00-9.00)	NS
Umbilical artery pH, median (IQR)	7.316 (7.289-7.343)	7.293 (7.258- 7.325)	<.05
CD	17.1% (7/41)	51.6% (16/31)	<.01

19 BMI, Body mass index; CD, Chronic deciduitis; HDP, Hypertensive disorders of  
 20 pregnancy; IVF, *In vitro* fertilization; LFD, Light-for-date infant; NS, Not significant.

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