

# Antioxidants and Therapeutic Targets in Ovarian Clear Cell Carcinoma.

著者	AMANO Tsukuru, MURAKAMI Atsushi, MURAKAMI Takashi, CHANO Tokuhiko
journal or publication title	Antioxidants (Basel, Switzerland)
volume	10
number	2
page range	187
year	2021-01-28
URL	<a href="http://hdl.handle.net/10422/00012907">http://hdl.handle.net/10422/00012907</a>

doi: 10.3390/antiox10020187(<https://doi.org/10.3390/antiox10020187>)



Review

# Antioxidants and Therapeutic Targets in Ovarian Clear Cell Carcinoma

Tsukuru Amano <sup>1,\*</sup>, Atsushi Murakami <sup>1</sup>, Takashi Murakami <sup>1</sup> and Tokuhiro Chano <sup>2,\*</sup>

<sup>1</sup> Department of Obstetrics & Gynecology, Shiga University of Medical Science, Tsukinowa-cho, Seta, Otsu, Shiga 520-2192, Japan; atsushim@belle.shiga-med.ac.jp (A.M.); tm@belle.shiga-med.ac.jp (T.M.)

<sup>2</sup> Department of Clinical Laboratory Medicine and Medical Genetics, Shiga University of Medical Science, Tsukinowa-cho, Seta, Otsu, Shiga 520-2192, Japan

\* Correspondence: tsukuru@belle.shiga-med.ac.jp (T.A.); chano@belle.shiga-med.ac.jp (T.C.); Tel.: +81-77-548-2267 (T.A.); +81-77-548-2621 (T.C.)

**Abstract:** Ovarian clear cell carcinomas (OCCCs) are resistant to conventional anti-cancer drugs; moreover, the prognoses of advanced or recurrent patients are extremely poor. OCCCs often arise from endometriosis associated with strong oxidative stress. Of note, the stress involved in OCCCs can be divided into the following two categories: (a) carcinogenesis from endometriosis to OCCC and (b) factors related to treatment after carcinogenesis. Antioxidants can reduce the risk of OCCC formation by quenching reactive oxygen species (ROS); however, the oxidant stress-tolerant properties assist in the survival of OCCC cells when the malignant transformation has already occurred. Moreover, the acquisition of oxidative stress resistance is also involved in the cancer stemness of OCCC. This review summarizes the recent advances in the process and prevention of carcinogenesis, the characteristic nature of tumors, and the treatment of post-refractory OCCCs, which are highly linked to oxidative stress. Although therapeutic approaches should still be improved against OCCCs, multi-combinatorial treatments including nucleic acid-based drugs directed to the transcriptional profile of each OCCC are expected to improve the outcomes of patients.

**Keywords:** ovarian clear cell carcinoma; endometriosis; antioxidant; cancer stemness



**Citation:** Amano, T.; Murakami, A.; Murakami, T.; Chano, T. Antioxidants and Therapeutic Targets in Ovarian Clear Cell Carcinoma. *Antioxidants* **2021**, *10*, 187. <https://doi.org/antiox10020187>

Academic Editor:

Jose Manuel Martinez-Martos

Received: 30 December 2020

Accepted: 24 January 2021

Published: 28 January 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Ovarian cancer is the eighth most common cancer affecting women worldwide, with an estimated 295,000 new cases in 2018 [1] and has a high mortality rate. In 2018 alone, approximately 185,000, 14,000, and 4800 deaths were reported worldwide, in the United States and Japan due to ovarian cancer, respectively [2,3]. Ovarian cancers can be classified into the following five pathological types: high-grade serous carcinomas (HGSCs), low-grade serous carcinomas (LGSCs), mucinous carcinomas (MCs), endometrioid carcinomas (ECs), and clear cell carcinomas (CCCs). The percentages of each pathological type among all ovarian cancers in the United States and Japan are as follows: serous carcinomas (combining HGSCs and LGSCs) 70%/36%, MCs 11%/11%, CCCs 5%/24%, and ECs 10%/17%, respectively. CCCs and ECs are much more common and great concerns in Japan and East Asian nations than in Europe or the United States [4,5]. HGSCs, which account for most of the serous carcinomas, are generally sensitive to chemotherapy. Furthermore, many studies have been conducted, providing new information. *TP53* mutations are detected in the majority of cases, and homologous recombination deficiency (HRD), including *BRCA* inactivation, have also been reported in approximately 50% of cases [6,7]. In this regard, novel treatments, such as the application of poly ADP-ribose polymerase (PARP) inhibitors, are being developed for HGSCs. LGSCs and MCs are relatively rare. LGSCs account for only 5–10% of ovarian serous carcinoma, and the majority of ovarian MCs are considered metastatic tumors derived from the gastrointestinal tract, while only  $\leq 3\%$  of MCs are regarded to be truly originating from the ovary [8,9].

Ovarian ECs and ovarian clear cell carcinomas (OCCCs) are both known as endometriosis-associated ovarian cancers, and are similar in that they are often associated with genetic mutations such as *ARID1A* and *PIC3CA*. Differences between these two are also apparent. ECs display estrogen and progesterone receptors [10] and are not significantly associated with antioxidant molecules such as hepatocyte nuclear factor 1 homeobox B (HNF1B) or mitochondrial superoxide dismutase (SOD2). The prognosis of ECs is relatively good, because they are sensitive to chemotherapy [11]. By contrast, OCCCs rarely express estrogen or progesterone receptors, and often overexpress HNF1B and SOD2. As overexpression of HNF1B and SOD2 confers resistance to oxidative stress in cancer cells, OCCCs possess strong resistance to oxidative stress caused by cancer treatments such as chemotherapy. Therefore, conventional anti-cancer drugs are ineffective against OCCCs, which often progress to advanced stages, and prognosis is extremely poor [12–14]. OCCC frequency is especially high in East Asian countries including Japan, and no effective treatment exists for OCCCs. As such, studies of molecular signatures of this type of cancer, and establishment of novel treatment methods, are urgently needed.

Among the various pathological types of ovarian cancers, OCCCs are most strongly linked to oxidative stress tolerance. Oxidative stress causes cancer by directly damaging DNA. Recently, it has been elucidated that molecular abnormalities involved in oxidative stress generation are deeply involved not only in carcinogenesis but also in cancer progression, such as cancer invasion and metastasis. The involvement of oxidative stress in OCCCs can be divided into the following two categories: (a) carcinogenesis from endometriosis to OCCC and (b) factors related to treatment after carcinogenesis.

This review focuses on the relationship between OCCCs and oxidative stress, the process and prevention of carcinogenesis, the nature of tumors, and treatment after carcinogenesis.

## 2. Linking Oxidative Stress and Carcinogenesis from Endometriosis to OCCCs

A strong association between epithelial ovarian cancer and endometriosis, a common gynecologic disorder affecting approximately 10% of reproductive-age women [15], has long been suggested. Among epithelial ovarian cancers, OCCCs are considered the most closely associated with endometriosis. A pooled analysis of case-control studies indicated that a self-reported history of endometriosis is associated with a higher increased risk of OCCCs (odds ratio: 3.05) than the other histologic subtypes of ovarian cancers [16]. Additionally, the coexistent rate of endometriosis in the same ovary in each histologic subtype of ovarian cancer is reported to be 35.9% in OCCCs, 19% in ECs, 4.5% in SCs, and 1.4% in MCs [17].

Endometriosis contributes to the carcinogenesis of epithelial ovarian cancers through chronic inflammation, local hyperestrogenism, and oxidative stress. As represented by colitis-associated colorectal cancer, chronic inflammation has long been known to cause malignant transformation of cells and carcinogenesis [18,19]. Several inflammatory mediators, such as TNF- $\alpha$ , IL-6, and TGF- $\beta$ , have been shown to participate in both the initiation and progression of endometriosis-associated ovarian cancers [20]. The abundance of estradiol in endometriotic lesions is caused by the locally increased expression of aromatase and steroidogenic acute regulatory protein combined with the decreased expression of 17 $\beta$ -hydroxysteroid dehydrogenase 2 [21]. These hormonal pathways are thought to be more closely associated with hormonal receptor-positive ovarian endometrioid carcinomas. On the other hand, as gene abnormalities associated with oxidative stress response and reactive oxygen species (ROS) metabolism are often detected [22], oxidative stress is considered to be most involved in the carcinogenesis of endometriosis into OCCCs. Endometriosis often results in chocolate (endometriotic) cyst formation, containing old blood with excess iron in the ovary. Iron and its metabolites contribute to the generation of ROS through the Fenton reaction, acting as inducers of DNA damage and sugar, lipid, and protein modifications, leading to carcinogenesis [23,24]. The carcinogenicity of iron compounds has been clearly demonstrated in previous animal experiments. Interestingly, several studies have shown that renal clear-cell carcinomas, which often share molecular features with OCCCs,

were produced by intraperitoneal iron chelate injection [25,26]. Oxidative stress induced by excess heme production and iron accumulation could also be an important trigger in malignant transformation of endometriosis to OCCCs [27].

### 3. Attempts to Prevent OCCCs Developing from Endometriosis

In OCCCs, abnormalities are often found in genes associated with oxidative stress and ROS metabolism [22]. The elimination of persistent inflammation and ROS is important to prevent carcinogenesis from endometriosis to OCCC. Surgery is a useful tool to prevent endometriosis. A nested case-control study in Sweden revealed that compared to controls, a one-sided oophorectomy or radical extirpation of all visible endometriosis reduced the risk of later development of ovarian cancer to 19% and 30%, respectively. However, hormonal treatments, such as combined oral contraceptives, gestagens (including oral drugs or levonorgestrel-containing intrauterine devices), danazol, and gonadotropin-releasing hormone agonists, did not mitigate cancer risk [28]. Several cases of OCCCs arising from endometrioma during hormonal treatment have also been reported [29]. On the other hand, several studies reported that oral contraception reduces the risk of ovarian cancer among women with and without endometriosis by suppressing ovulation [30,31]. Through the collaborative analysis of data from 45 epidemiological studies, Basel et al. reported that after 5 years of oral contraceptive use, the risk of OCCC was reduced by 21.3% [31]. However, a few study limitations are attributed to the retrospective and contain data from areas with a low frequency of OCCCs among the population. Hormonal therapy using low-dose estrogen-progestin or dienogest (progestin medication) may suppress the progression of endometriosis to OCCC. A large-scale prospective study by the Japan Endometrioma Malignant-transformation Study (JEMS) is currently underway in Japan, where OCCCs are frequent in the population. Moreover, further research is needed to clarify this point.

Since oxidative stress is involved in the formation of OCCCs, antioxidant intake may be effective in preventing its development. Vitamin A (carotenoid), C, E, flavonoids, and isothiocyanate are known antioxidant supplements [32–35]. Thus, diets rich in vegetables and fruits, which are good sources of antioxidants, are considered healthy. Antioxidants may prevent or delay various steps associated with carcinogenesis [36–38]. Carotenoid astaxanthin reduces oxidative stress, and inflammation [39,40] exerts a highly protective antioxidant effect [41]. Astaxanthin has been shown to decrease DNA damage and improve the immune response in healthy women after 8 weeks of intake [42]. In both in vitro and in vivo experiments, the use of astaxanthin significantly inhibited tumor formation and growth and exhibited anticancer properties [43–46]. Flavonoids inhibit multiple enzymes involved in cancer cell growth and arrest the cell cycle and tumor regression by activating the mitochondrial pathway of apoptosis [47,48]. Isothiocyanate inhibits the growth of ovarian cancer cells by inducing apoptosis in in vitro experiments [49]. Animal experiments have demonstrated that isothiocyanate exerts inhibitory effects on the carcinogenesis of both forestomach and lung cancers induced by the carcinogen benzopyrene [50].

As an epidemiological investigation, the effects of the daily intake of antioxidant supplements on ovarian cancer were investigated in multiple population-based case-control and prospective cohort studies, and several meta-analyses have been published. These studies indicated that the intake of dietary vitamins C, D, E, and isothiocyanate are not associated with the risk of ovarian cancers [51–53]. On the other hand, a higher dietary intake of vitamin A, including carotenoids and flavonoids, may lower the risk of ovarian cancer [54–56] (Table 1). In these studies, the subjects were members of the general population and did not necessarily have increased levels of oxidative stress. Antioxidant intake may be more effective in preventing carcinogenesis in people with endometriosis, as endometriosis treatment reduces the risk of developing ovarian cancers later. Because OCCC carcinogenesis is particularly affected by oxidative stress, antioxidants may be effective for its prevention. However, it should be noted that these studies did not consider the pathological type of ovarian cancer; thus, whether this leads to the prevention of

OCCCs remains unknown. Furthermore, it must also be taken into account that the intake of vitamin A and beta-carotene has a negative effect on other carcinomas [54].

**Table 1.** Antioxidant supplements and foods against ovarian cancers.

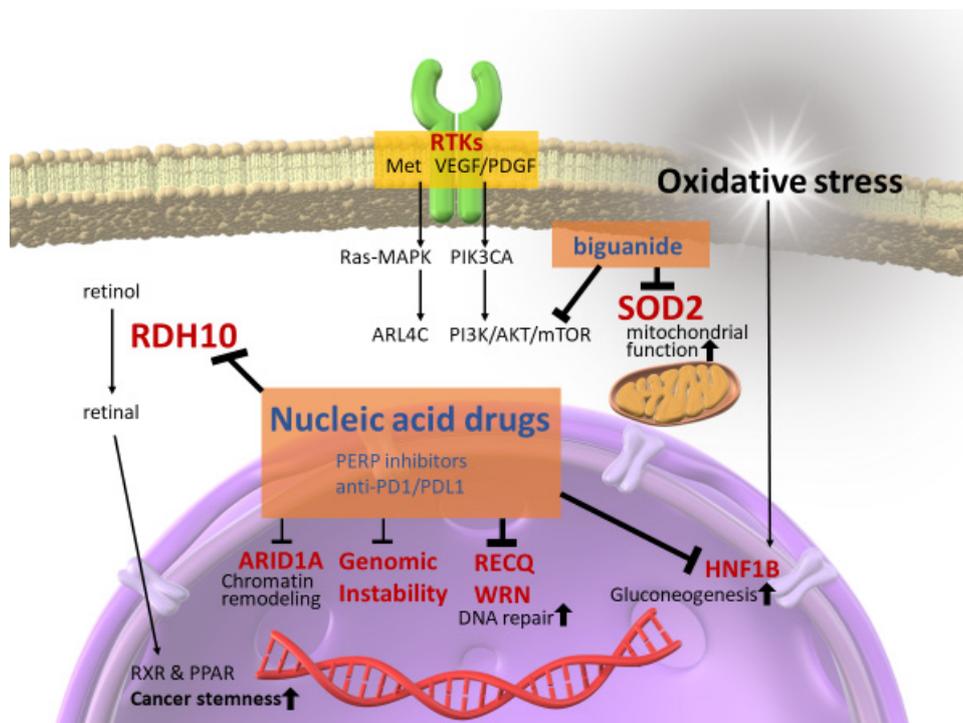
Clinical Research	Antioxidant	Result	Reference
Meta-analysis	Vitamin A	Reduced the risk of ovarian cancer.	[52]
Meta-analysis	Vitamin C	No significant effect on the risk of ovarian cancer.	[53]
Systematic review	Vitamin D	No significant effect on the risk of ovarian cancer.	[55]
Meta-analysis	Vitamin E	No significant effect on the risk of ovarian cancer.	[51]
Meta-analysis	Flavonoid	Reduced the risk of ovarian cancer.	[56]
Cohort study	Flavonoid	Reduced the risk of ovarian cancer.	[57]
	Isothiocyanate	No significant effect on the risk of ovarian cancer.	

A recent systematic review, cohort study, and meta-analyses indicated that vitamin A and flavonoid intake might reduce the risk of ovarian cancers, although vitamins C, D, E, and isothiocyanate have no significant effects on the risks. The preventive effects of flavonoids and vitamin A on OCCC (ovarian clear cell carcinoma) remain unclarified.

Currently, surgery is suitable to prevent carcinogenesis from chocolate cysts. However, the effects of hormonal therapies and dietary additives consisting of strong antioxidants, flavonoids, and isothiocyanates will require further investigation.

#### 4. Molecular Characteristics in OCCCs Related to Anti-Oxidative Pathway

As mentioned above, in OCCCs, which arise from endometriosis under massive oxidative stress, abnormalities are often detected in genes associated with the oxidative stress response and ROS metabolism [22]. Several antioxidant molecules are involved in OCCC carcinogenesis. Among them, the overexpression of hepatocyte nuclear factor 1 homeobox B (HNF1B), a major homeobox-containing protein, also known as transcription factor-2, is highly important. Under hypoxia and acidosis, HNF1B can modify and adapt cancer cells to survive through a process between gluconeogenesis and glycolysis, commonly known as the Warburg effect [58]. Tsuchiya et al. [14] first reported HNF1B overexpression in OCCCs and showed that reduced HNF1B expression considerably increased the apoptosis rate in two OCCC cell lines. Overexpression of HNF1B was observed in endometrial tissues adjacent to OCCC tumors, suggesting that HNF1B overexpression is an early event in OCCC carcinogenesis. Kato et al. [59] found that hypomethylation of the CpG island of HNF1B induced its overexpression in OCCCs, indicating that overexpression in OCCC was also caused by epigenetic changes rather than by mutations. Moreover, recent research has revealed that HNF1B promotes the dedifferentiation of cancer stem-like cells (CSCs) via activation of the Notch pathway and enhancing the invasive potential and epithelial-mesenchymal transition in cancer cells [60]. Anti-oxidative pathways are deeply involved in carcinogenesis and therapeutic resistance in OCCCs. As oxidative stress tolerance represents therapeutic resistance, OCCCs usually exhibit poor and fatal prognoses, even during gradual progression. OCCC has low sensitivity to platinum- and taxane-based chemotherapy. Therefore, the prognosis of OCCCs is extremely poor, particularly in its advanced stages [61,62]. Previous studies have revealed the role of HNF1B in driving the expression of several characteristic genes associated with OCCCs [63], stimulating metabolic changes to promote gluconeogenesis, glycogen accumulation, and aerobic glycolysis [64], inducing chemotherapeutic resistance by suppressing sulfatase-1 (Sulf-1), an extracellular sulfatase catalyzing the 6-O desulfation of heparan sulfate glycosaminoglycans [65], and reducing the activity of immunological checkpoints against tumors. Thus, HNF1B plays an important role in therapeutic resistance via oxidative stress tolerance in OCCCs (Figure 1).



**Figure 1.** Activating pathways and targeting proposals in ovarian clear cell carcinomas. In ovarian clear cell carcinomas, downstream of receptor tyrosine kinases (RTKs), AT-rich interactive domain 1A (*ARID1A*)-related chromatin remodeling factors, and genomic instability, including MSI-H, are activated. These are currently being targeted. However, other therapeutic strategies, such as nucleic acid-based drugs, RDH10, RECQL1, WRN, and HNF1B, should be targeted in the future to reduce cancer stemness, induce cancer-specific synthetic lethality, and reduce gluconeogenesis, together with a drug repositioning strategy against SOD2 anti-oxidative stress molecules.

Mitochondrial superoxide dismutase (SOD2) is an antioxidant enzyme that metabolizes superoxide in mitochondria and plays an important role in maintaining mitochondrial function through oxidative stress tolerance. SOD2 is highly expressed in the ectopic endometrium compared to normal endometrium, promoting cell proliferation and migration in ovarian endometriosis [66]. SOD2 is also highly expressed in OCCCs, and its oxidative stress tolerance appears to contribute to carcinogenesis [67,68]. SOD2 overexpression also promotes tumor growth and metastasis in OCCCs. Hemachandra et al. [67] found that SOD2 was more highly expressed in OCCCs than in any other epithelial ovarian cancer subtypes, and its overexpression contributes to tumor growth and metastasis in a chorioallantoic membrane model. The study also indicated that SOD2 expression was associated with increased cell proliferation, migration, outgrowth on collagen, spheroid attachment, and Akt phosphorylation in ES-2 OCCC cells. Therefore, SOD2 is regarded as a pro-tumorigenic or metastatic factor in OCCCs. Clinical studies have also demonstrated that high SOD2 expression was observed in 76% (33 out of 41) of OCCCs, and SOD2 overexpression was correlated with poor prognoses for OCCCs [68]. Accordingly, SOD2 is considered to be involved in therapeutic refraction through oxidative stress resistance in OCCCs (Figure 1).

## 5. Oxidative Stress and Cancer Stemness of OCCC

Recently, CSCs resistant to oxidative stress have been associated with the recurrence and metastasis of malignant tumors [69]. In other words, CSCs can survive severe oxidative stress induced by radiation therapy or chemotherapy and contribute to recurrence and metastasis. Because OCCC is often refractory to chemotherapy or relapse even after remission, it is suggested that CSCs are involved in the recurrence or metastasis of OCCC. High expression levels of aldehyde dehydrogenase 1 (ALDH1), a CSC marker, and Nrf2,

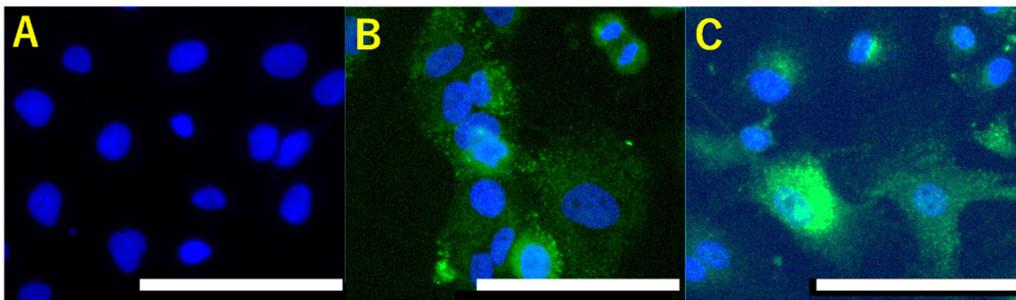
a key transcriptional factor of the antioxidant system, were both associated with a poor prognosis in OCCC [70]. Furthermore, our group recently found that retinol dehydrogenase 10 (RDH10), enzymes related to vitamin A metabolism and gluconeogenesis, can reflect cancer stemness through precise analyses of the RAB39A (a member of the RAS oncogene family)-RXRB (retinoid X receptor beta) axis. The RAB39A-RXRB axis drives cancer stemness and tumorigenesis; consequently, the downregulation of this pathway leads to poor sphere formation and xenotransplantable function in several types of malignancies, such as sarcomas, adrenal, lymphoid, and testicular tumors [71]. On the other hand, some subpopulations of cancer cells could produce vividly growing spheres regardless of RAB39A repression. Therefore, under continuous RAB39A repression, we compared vividly growing cancer spheres to poor ones via RNA-seq transcriptional analysis, whose original sequencing data have been deposited in DDBJ/EMBL/GenBank (accession no. DRA010748). As a result, vividly growing cancer spheres were found to be significantly related to the upregulation of 79 genes (Supplementary Table S1) and of signaling pathways on retinoic acid (RA), vitamin A, and carotenoid metabolism, which were listed in statistically high orders of ranking (Table 2). In the pathway contributing to nuclear RXRB function (WP716\_83589), retinol dehydrogenase 10 (RDH10), which converts retinol to all-trans RA and is indispensable for RA synthesis as the predominant enzyme [72,73], was focused on as a member of the 79 genes.

**Table 2.** Upregulated pathways in vividly growing cancer spheres.

Pathway	<i>p</i> -Value (RNAseq from Vividly Growing Cancer Spheres)
Hs_Integrated_Cancer_Pathway_WP1971_82939	$1.02 \times 10^4$
Hs_Intrinsic_Pathway_for_Apoptosis_WP1841_83332	0.002799811
Hs_Signaling_by_Retinoic_Acid_WP3323_83286	0.003082763
Hs_Vitamin_A_and_Carotenoid_Metabolism_WP716_83589	0.003229062
Hs_BMAL1-CLOCK, NPAS2_activates_circadian_gene_expression_WP3355_83343	0.003687094
Hs_Integrated_Breast_Cancer_Pathway_WP1984_82941	0.004416806
Hs_Pre-NOTCH_Expression_and_Processing_WP2786_83418	0.004513508
Hs_Lipid_storage_and_perilipins_in_skeletal_muscle_WP2887_85092	0.011662396
Hs_Uptake_and_function_of_anthrax_toxins_WP3390_83389	0.013593029
Hs_miRNA_Regulation_of_DNA_Damage_Response_WP1530_84694	0.013785134

Pathways were explicitly upregulated in vividly growing cancer spheres, irrespective of continuous RAB39A repression. Note: The pathway of retinoic acid signaling, namely vitamin A and carotenoid metabolism, is listed in the superior order of the top 10 rankings.

Interestingly, RDH10 is involved in insulin signaling and contributes to gluconeogenesis via conversion from retinal to all-trans RA [74]. Gluconeogenesis and carbon hydrate storage are characteristics of OCCC phenotypes. Irrespective of RAB39A downregulation, RDH10 overexpression can result in the continuous activation of nuclear RXRB function and connect to CSC and carbon hydrate storage characteristics. OCCC cell lines specifically express abundant RDH10, rather than other types of ovarian cancer cells (Figure 2). RA is known to suppress cancer stemness and tumorigenesis because RA promotes cell differentiation, cell cycle arrest, and apoptosis via the heterodimer of retinoic acid receptor (RAR) and retinoid X receptor (RXR) [75–77]. In contrast, Schung et al. demonstrated that RA promotes cell survival in fatty acid-binding protein 5 (FABP5) cells through peroxisome proliferator-activated receptor beta (PPAR $\beta/\delta$ ) [78]. Interestingly, the overexpression of FABP5 is an unfavorable prognostic marker in renal clear-cell carcinoma, which shows pathological similarities to OCCC [79]. Furthermore, RDH10 overexpression promotes tumor cell proliferation and correlates with patient survival time in gliomas [80].



**Figure 2.** Abundant retinol dehydrogenase 10 (RDH10) expression in ovarian clear cell carcinomas. In contrast to SK-OV-3, ovarian serous cell carcinoma (A), ovarian clear cell carcinoma cells, OVISE (B), and TOV-21 (C) express high levels of retinol dehydrogenase 10 (RDH10). Scale bars: 100  $\mu$ m.

Overall, RDH10 overexpression may imply cancer stemness and tumorigenesis in OCCC, resulting in a difficult prognosis refractory to existing therapies. RDH10 may serve as a novel diagnostic and therapeutic target for OCCC.

## 6. Therapeutic Targets for OCCCs in the Present and Future

As mentioned above, conventional standard treatments are less effective for OCCCs because of their strong tolerance to oxidative stress. Thus, to overcome the therapeutic difficulties associated with ovarian cancers, especially for OCCCs, novel therapeutics for recurrent or refractory cases are urgently needed. At present, several molecular targets have been proposed for OCCCs, which are categorized into the following groups: pathways related to receptor tyrosine kinases (RTKs), AT-rich interactive domain 1A (ARID1A)-related chromatin remodeling factors, and molecules associated with immune checkpoints. Some clinical trials have already been completed or are currently conducting, and we have summarized them (Table 3).

RTK receptors are located on the cell surface and play an important role in regulating cell proliferation, differentiation, survival, metabolism, and migration. Both the phosphoinositide 3-kinase/AKT/mammalian target of rapamycin (PI3K/AKT/mTOR) and epidermal growth factor/Ras/mitogen-activated protein kinase (EGF/Ras/MAPK) pathways are downstream pathways of RTKs. Mutations in phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha (*PIK3CA*), the loss of phosphatase and tensin homolog (*PTEN*), the amplification of human epidermal growth factor receptor 2 (*HER2*), overexpression of *MET* (also known as hepatocyte growth factor receptor; *HGFR*), and ADP-ribosylation factor-like 4C (*ARL4C*) have been shown to activate these pathways in OCCCs [10,81–83]. In several studies, the inhibition of these molecules has shown the potential to suppress OCCCs. For example, *MET* inhibitors significantly decreased the proliferation and increased the apoptosis of OCCC cells in vitro, and suppressed tumor growth in xenograft models of OCCC in vivo [82]. Despite its effectiveness in vitro, no clinical advantages have been observed for inhibitors of downstream pathways of RTKs in treating OCCCs. The *MET* inhibitor cabozantinib was clinically ineffective in treating 13 patients with recurrent OCCCs [84]. The combination of temsirolimus and carboplatin or paclitaxel was also investigated in patients with advanced OCCCs. However, compared to conventional treatments, this regimen did not significantly increase the rate of progression-free survival [85]. Sunitinib, another RTK targeting inhibitor of VEGF and PDGF signaling, demonstrated minimal activity in second- and third-line treatments of persistent or recurrent OCCCs [86]. Thus far, no RTK inhibitors have demonstrated efficacy against OCCC in clinical trials. Further studies are needed to identify more effective drugs combined with PI3K/AKT/mTOR inhibitors and the mutations associated with OCCC that can be targeted by PI3K/AKT/mTOR inhibitors.

**Table 3.** Molecular targeting drugs and the clinical trials to treat OCCCs (ovarian clear cell carcinomas).

Category	Target Molecules	Clinical Research	Result	Reference
RTKs and related molecules	MET	already completed	minimal activity	[84]
	PI3K/AKT/mTOR	already completed	clinically ineffective	[85]
	VEGFR/PDGFR	already completed	minimal activity	[86]
	VEGFR/PDGFR/FGFR	Currently conducting	—	NCT02866370
ARID1A chromatin remodeling factor	EZH2/glutathione	—	—	[87–89]
Immune checkpoint proteins	PD-L1	Currently conducting	—	NCT03405454
	PD-1/CTLA-4	Currently conducting	—	NCT03355976
	TIM-1	Currently conducting	—	NCT02837991

Clinical trials have been completed for several inhibitors of RTK-related pathways, but no significant effect against OCCCs has been found. Multiple clinical trials are currently underway to evaluate immune checkpoint inhibitors in OCCC.

ARID1A chromatin remodeling abnormalities are also useful therapeutic targets for OCCC [13]. The *ARID1A* gene encodes BAF250a, a subunit of the switch/sucrose non-fermentable (SWI/SNF) chromatin remodeling complex modifies the structure of chromatin via histone octamer ejection, octamer sliding, or local chromatin unwrapping to allow for the binding of other transcription factors [13]. Additionally, mutations in *ARID1A* contribute to AKT phosphorylation and induce PI3K pathway activation [90]. In *ARID1A*-mutated OCCCs, inhibition of the enhancer of zeste homolog 2 (EZH2) histone methyltransferase activity could induce synthetic lethality, including the suppression of PI3K/AKT signaling [87]. Additionally, Berns et al. [88] recently found that small-molecule inhibitors of the bromodomain and extra-terminal domain (BET) family of proteins inhibit the proliferation of *ARID1A*-mutated OCCC cells by reducing the expression of multiple SWI/SNF members in vitro and in vivo. Furthermore, recent research has demonstrated that *ARID1A*-deficient cancer cells have low levels of glutathione due to the decreased expression of SLC7A11 and are specifically vulnerable to the inhibition of the antioxidant glutathione and glutamate-cysteine ligase synthetase catalytic subunit, a rate-limiting enzyme for glutathione synthesis [89]. APR-246, a glutathione inhibitor, can act as an effective agent to induce synthetic lethality in *ARID1A*-deficient cancer cells [89]. Thus, EZH2, BET, and APR-246 are promising new drugs for the treatment of *ARID1A*-mutated OCCCs.

In recent years, significant progress has been made in targeting therapy to immune checkpoint proteins in cancers. In particular, immune checkpoint inhibitors (ICIs) by anti-programmed death receptor-1/programmed death-ligand 1 (PD-1/PD-L1) antibodies for high microsatellite instability (MSI-H) and high tumor mutation burden (TMB-H) tumors are effective in multiple studies [91–94]. The objective response rate and progression-free survival rate by PD-1 blockade were reported to be 40% and 78% for mismatch repair-deficient colorectal cancers and 0% and 11% for mismatch repair-proficient colorectal cancers, respectively [92]. Moreover, immunotherapy with PD-1 blockade for patients with advanced mismatch repair-deficient cancers across 12 different types achieved a 53% objective response rate and 21% complete response rate [91]. Furthermore, recent studies suggest that TMB can also predict the response to PD-1 inhibitors [93] and is associated with improved survival in patients receiving ICI across various cancer types [94].

As for OCCCs, recent research demonstrated that about 7% (4 out of 57) of OCCC cases had MSI-H cancers without any MMR mutations [95]. Feinberg reported that despite the presence of only four (1.6%) MSI-H tumors in 254 OCCC cases, 23 (9.0%) tumors with high TMB were found [96]. Additionally, it was also revealed that *ARID1A* alterations were associated with high TMB levels across cancer types and may cooperate with ICI treatment [97]. However, the relationship between *ARID1A*-mutated OCCC and efficacy of ICI treatment remains unknown. In summary, approximately 10% of OCCCs carry properties of MSI-H or TMB-H and may benefit from ICIs. For some OCCC patients, immunotherapy with anti-PD-1/PD-L1 antibodies has a high potential to be used as an effective treatment strategy. Multiple clinical trials are currently underway to evaluate PD-

1/PD-L1 inhibitors in OCCCs. In tumor immunotherapy, another therapeutic candidate is T-cell immunoglobulin and mucin domain protein 1 (TIM-1), which regulates immune responses on human T cell surfaces. TIM-1, expressed on a high percentage of OCCC cells, exhausts T cell immunity in cancer microenvironments [98]. Clinical trials of CDX-014, an anti-TIM-1 antibody covalently linked to the potent cytotoxin, monomethyl auristatin E (MMAE), are currently being conducted in OCCC patients. Further studies of targeted therapies for immune checkpoint proteins are eagerly awaited.

As mentioned under the headings “Molecular characteristics in OCCCs related to anti-oxidative pathway” and “Oxidative stress and cancer stemness of OCCC,” molecules conferring oxidative stress resistance, including HNF1B, SOD2, and RDH10, are deeply implicated in therapeutic refractivity. Together with SOD2, HNF1B and RDH10 are major potential targets for OCCC treatment. However, molecular inhibitors of the latter two molecules have not been identified, and difficulties have arisen in development of chemical agents that inhibit them. In cancer cell subpopulations, especially in CSCs, the mitochondrial respiratory chain relies heavily on bioenergetic and biosynthetic processes. Therefore, mitochondrial function can be a target for therapeutic strategies in several cancers [86,99–103]. Since SOD2 plays an important role in maintaining mitochondrial function through oxidative stress tolerance, drugs suppressing mitochondrial function should be effective in treating SOD2-abundant OCCC. Replacement therapy or drug repositioning using biguanides, agents for treating diabetes mellitus that inhibit complex 1 of the mitochondrial respiratory chain may target tumor cell mitochondria and thereby improve the therapeutic effect on OCCCs [104]. It has been confirmed that metformin, commonly used as a first-line biguanide treatment for type 2 diabetes, can selectively inhibit mitochondrial respiratory chain complex 1 in various cancer cell lines [105–108]. As metformin is not metabolized, but almost all are excreted by the kidney, the half-life of plasma levels is long. Therefore, metformin accumulates in organs, and tissue concentration is considered to be higher than that of plasma [107]. Furthermore, many studies have shown that metformin accumulates at high concentrations in the mitochondria [107,109]. In addition to mitochondrial inhibition, metformin also demonstrates an anti-tumor effect through several routes, including immune-mediated, mammalian target of rapamycin (m-TOR), and AMP-activated protein kinase (AMPK) [110]. The follow-up program post-surgical resection of OCCC may involve drug repositioning using metformin; however, this will need to be verified by clinical cohort studies in the future. In addition, Molina et al. recently discovered IACS-010759, a new clinical-grade small-molecule inhibitor of complex I of the mitochondrial electron transport chain, which inhibits tumor growth in models of brain cancer and AML at well-tolerated doses in vivo [103]. SOD2-abundant OCCC may also be an adequate candidate for this new inhibitor.

Targeting DNA helicases is another therapeutic strategy for MSI-H tumors, which may be suitable for use in combination with immune checkpoint inhibitors for cancer chemotherapy. Human DNA helicases RECQL1 and WRN proteins have been reported as therapeutic targets for several cancers, including ovarian cancers [111,112]. As for OCCCs, 9 out of 21 (43%) clinical cases showed high levels of RECQL1 expression by immunohistopathology, and RECQL1-siRNA significantly inhibited the proliferation of OCCC cell lines [112]. Importantly, recent research revealed that WRN induced double-stranded DNA breaks and promoted apoptosis and cell cycle arrest selectively in MSI-H cancer models [113]. In other words, WRN was selectively essential for MSI-H cancers, and the inhibition of WRN-induced synthetic lethality in MSI-H cancer models. These findings indicate that at least some OCCC patients benefit from RECQL1 or WRN inhibition.

In the future, further therapeutic options will need to be developed to improve OCCC achievements. In addition to drug repositioning against SOD2 anti-oxidative stress molecules and targeting RECQL1 and WRN to induce cancer-specific synthetic lethality, it is advisable to implement nucleic acid-based drugs, such as siRNA and antisense oligonucleotides, to treat OCCCs effectively and to adapt them to the transcriptional profile of individual tumor characteristics in the era of precision medicine. Integrative in silico

approaches have indicated that only 10–15% of human proteins are druggable and that only 10–15% of human proteins are disease-modifying [114]. Hence, developing chemical agents that inhibit RDH10, HNF1B, and ARID1A is a challenge. However, considering the implementation of nucleic acid-based drugs, RDH10, HNF1B, ARID1A, RECQ, and WRN helicases should be included in future therapeutic targets.

## 7. Conclusions

In this review, we have briefly summarized recent advances in the process and prevention of carcinogenesis, the characteristic nature of tumors, and the treatment of post-refractory OCCCs, which are highly linked to oxidative stress. The removal of oxidative stress suppresses the development of OCCCs in endometriosis. Strong antioxidants, such as vitamin A, carotenoids, or flavonoids, may help prevent carcinogenesis of OCCCs. However, the stress tolerance properties of OCCCs induce therapeutic resistance, making their treatment difficult. Antioxidants display bidirectional effects toward endometriosis and OCCCs. Elimination of oxidative stress, including by uptake of antioxidants, is highly effective in preventing progression from endometriosis to OCCCs, but, antioxidants are not suitable for treatment of established OCCCs, in which oxidative stress tolerance has accrued, providing therapeutic resistance. In OCCC therapeutics, inhibition of oxidative stress tolerance molecules is essential. The genetic and biological characteristics of OCCCs are being gradually evaluated, and the therapeutic effects of various anti-cancer drugs, molecular targeting drugs, drug repositioning strategies, and immunotherapies are being verified. Further studies will be needed to identify novel molecular targets, and studies on precision medicine, combining multiple treatments based on the genetic and molecular characteristics of individual tumors, will need to be conducted. Since the development of small molecular inhibitors for some undruggable molecules remains a challenge, it is essential that therapeutic approaches against OCCCs be improved, and that nucleic acid-based drugs and multi-combinatorial treatments corresponding to the transcriptional profile of each tumor be implemented.

**Supplementary Materials:** The following are available online at <https://www.mdpi.com/2076-3921/10/2/187/s1>, Table S1: Upregulated 79 genes in vividly growing cancer spheres.

**Author Contributions:** T.A.: Writing—Original Draft, A.M.: Investigation, T.M.: Supervision, T.C.: Writing—Review & Editing, Project administration. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Available online: <http://gco.iarc.fr/today/home> (accessed on 15 December 2020).
2. Available online: <https://www.cancer.org/research/cancer-facts-statistics/all-cancer-facts-figures/cancer-facts-figures-2018.html> (accessed on 15 December 2020).
3. Available online: [https://ganjoho.jp/reg\\_stat/statistics/stat/short\\_pred.html](https://ganjoho.jp/reg_stat/statistics/stat/short_pred.html) (accessed on 15 December 2020).
4. Yamagami, W.; Nagase, S.; Takahashi, F.; Ino, K.; Hachisuga, T.; Aoki, D.; Katabuchi, H. Clinical statistics of gynecologic cancers in Japan. *J. Gynecol. Oncol.* **2017**, *28*. [[CrossRef](#)] [[PubMed](#)]
5. Itamochi, H.; Kigawa, J.; Terakawa, N. Mechanisms of chemoresistance and poor prognosis in ovarian clear cell carcinoma. *Cancer Sci.* **2008**, *99*, 653–658. [[CrossRef](#)] [[PubMed](#)]
6. Shih, I.-M.; Kurman, R.J. Ovarian Tumorigenesis. *Am. J. Pathol.* **2004**, *164*, 1511–1518. [[CrossRef](#)]
7. Köbel, M.; Reuss, A.; Du Bois, A.; Kommoss, S.; Kommoss, F.; Gao, D.; Kalloger, S.E.; Huntsman, D.G.; Gilks, C.B. The biological and clinical value of p53 expression in pelvic high-grade serous carcinomas. *J. Pathol.* **2010**, *222*, 191–198. [[CrossRef](#)] [[PubMed](#)]
8. Kaldawy, A.; Segev, Y.; Lavie, O.; Auslender, R.; Sopik, V.; Narod, S.A. Low-grade serous ovarian cancer: A review. *Gynecol. Oncol.* **2016**, *143*, 433–438. [[CrossRef](#)] [[PubMed](#)]
9. Seidman, J.D.; Kurman, R.J.; Ronnett, B.M. Primary and Metastatic Mucinous Adenocarcinomas in the Ovaries. *Am. J. Surg. Pathol.* **2003**, *27*, 985–993. [[CrossRef](#)] [[PubMed](#)]
10. Köbel, M.; Kalloger, S.E.; Boyd, N.; McKinney, S.; Mehl, E.; Palmer, C.; Leung, S.; Bowen, N.J.; Ionescu, D.N.; Rajput, A.; et al. Ovarian Carcinoma Subtypes Are Different Diseases: Implications for Biomarker Studies. *PLoS Med.* **2008**, *5*, e232. [[CrossRef](#)]

11. Bouchard-Fortier, G.; Panzarella, T.; Rosen, B.; Chapman, W.; Gien, L.T. Endometrioid Carcinoma of the Ovary: Outcomes Compared to Serous Carcinoma After 10 Years of Follow-Up. *J. Obstet. Gynaecol. Can.* **2017**, *39*, 34–41. [[CrossRef](#)]
12. Kuo, K.-T.; Mao, T.-L.; Jones, S.; Veras, E.; Ayhan, A.; Wang, T.-L.; Glas, R.; Slamon, D.; Velculescu, V.E.; Kuman, R.J.; et al. Frequent Activating Mutations of PIK3CA in Ovarian Clear Cell Carcinoma. *Am. J. Pathol.* **2009**, *174*, 1597–1601. [[CrossRef](#)]
13. Jones, S.; Wang, T.-L.; Shih, I.-M.; Mao, T.-L.; Nakayama, K.; Roden, R.; Glas, R.; Slamon, D.; Diaz, L.A.; Vogelstein, B.; et al. Frequent Mutations of Chromatin Remodeling Gene ARID1A in Ovarian Clear Cell Carcinoma. *Science* **2010**, *330*, 228–231. [[CrossRef](#)]
14. Tsuchiya, A.; Sakamoto, M.; Yasuda, J.; Chuma, M.; Ohta, T.; Ohki, M.; Yasugi, T.; Taketani, Y.; Hirohashi, S. Expression Profiling in Ovarian Clear Cell Carcinoma. *Am. J. Pathol.* **2003**, *163*, 2503–2512. [[CrossRef](#)]
15. Zondervan, K.; Becker, C.M.; Missmer, S.A. Endometriosis. *N. Engl. J. Med.* **2020**, *382*, 1244–1256. [[CrossRef](#)] [[PubMed](#)]
16. Pearce, C.L.; Templeman, C.; Rossing, M.A.; Lee, A.; Near, A.M.; Webb, P.M.; Nagle, C.M.; Doherty, J.A.; Cushing-Haugen, K.L.; Wicklund, K.G.; et al. Association between endometriosis and risk of histological subtypes of ovarian cancer: Apooled analysis of case–control studies. *Lancet Oncol.* **2012**, *13*, 385–394. [[CrossRef](#)]
17. Nezhat, F.; Datta, M.S.; Hanson, V.; Pejovic, T.; Nezhat, C.; Nezhat, C. The relationship of endometriosis and ovarian malignancy: A review. *Fertil. Steril.* **2008**, *90*, 1559–1570. [[CrossRef](#)] [[PubMed](#)]
18. Landskron, G.; De La Fuente, M.; Thuwajit, P.; Thuwajit, C.; Hermoso, M.A. Chronic Inflammation and Cytokines in the Tumor Microenvironment. *J. Immunol. Res.* **2014**, *2014*, 149185. [[CrossRef](#)]
19. Balkwill, F.R.; Mantovani, A. Inflammation and cancer: Back to Virchow? *Lancet* **2001**, *357*, 539–545. [[CrossRef](#)]
20. Worley, J.M.J.; Welch, W.R.; Berkowitz, R.S.; Ng, S.-W. Endometriosis-Associated Ovarian Cancer: A Review of Pathogenesis. *Int. J. Mol. Sci.* **2013**, *14*, 5367–5379. [[CrossRef](#)]
21. Bulun, S.E. Endometriosis. *N. Engl. J. Med.* **2009**, *360*, 268–279. [[CrossRef](#)]
22. Yamaguchi, K.; Mandai, M.; Oura, T.; Matsumura, N.; Hamanishi, J.; Baba, T.; Matsui, S.; Murphy, S.K.; Konishi, I. Identification of an ovarian clear cell carcinoma gene signature that reflects inherent disease biology and the carcinogenic processes. *Oncogene* **2010**, *29*, 1741–1752. [[CrossRef](#)]
23. Toyokuni, S. Role of iron in carcinogenesis: Cancer as a ferrotoxic disease. *Cancer Sci.* **2009**, *100*, 9–16. [[CrossRef](#)]
24. Yamaguchi, K.; Mandai, M.; Toyokuni, S.; Hamanishi, J.; Higuchi, T.; Takakura, K.; Fujii, S. Contents of Endometriotic Cysts, Especially the High Concentration of Free Iron, Are a Possible Cause of Carcinogenesis in the Cysts through the Iron-Induced Persistent Oxidative Stress. *Clin. Cancer Res.* **2008**, *14*, 32–40. [[CrossRef](#)] [[PubMed](#)]
25. Li, J.L.; Okada, S.; Hamazaki, S.; Ebina, Y.; Midorikawa, O. Subacute nephrotoxicity and induction of renal cell carcinoma in mice treated with ferric nitrilotriacetate. *Cancer Res.* **1987**, *47*, 1867–1869. [[PubMed](#)]
26. Liu, M.; Okada, S. Induction of free radicals and tumors in the kidneys of Wistar rats by ferric ethylenediamine-N,N'-diacetate. *Carcinogenesis* **1994**, *15*, 2817–2821. [[CrossRef](#)] [[PubMed](#)]
27. Munksgaard, P.S.; Blaakaer, J. The association between endometriosis and ovarian cancer: A review of histological, genetic and molecular alterations. *Gynecol. Oncol.* **2012**, *124*, 164–169. [[CrossRef](#)] [[PubMed](#)]
28. Melin, A.-S.; Lundholm, C.; Malki, N.; Swahn, M.-L.; Sparén, P.; Bergqvist, A. Hormonal and surgical treatments for endometriosis and risk of epithelial ovarian cancer. *Acta Obstet. Gynecol. Scand.* **2013**, *92*, 546–554. [[CrossRef](#)] [[PubMed](#)]
29. Yoshino, O.; Minamisaka, T.; Ono, Y.; Tsuda, S.; Samejima, A.; Shima, T.; Nakashima, A.; Koga, K.; Osuga, Y.; Saito, S. Three cases of clear-cell adenocarcinoma arising from endometrioma during hormonal treatments. *J. Obstet. Gynaecol. Res.* **2018**, *44*, 1850–1858. [[CrossRef](#)]
30. Modugno, F.; Ness, R.B.; Allen, G.O.; Schildkraut, J.M.; Davis, F.G.; Goodman, M.T. Oral contraceptive use, reproductive history, and risk of epithelial ovarian cancer in women with and without endometriosis. *Am. J. Obstet. Gynecol.* **2004**, *191*, 733–740. [[CrossRef](#)]
31. Collaborative Group on Epidemiological Studies of Ovarian Cancer Ovarian cancer and oral contraceptives: Collaborative reanalysis of data from 45 epidemiological studies including 23,257 women with ovarian cancer and 87,303 controls. *Lancet* **2008**, *371*, 303–314. [[CrossRef](#)]
32. Nishida, Y.; Yamashita, E.; Miki, W. Quenching activities of common hydrophilic and lip-ophilic antioxidants against singlet oxygen using chemiluminescence detection System. *Carotenoid Sci.* **2007**, *11*, 16–20.
33. Martin, H.D.; Ruck, C.; Schmidt, M.; Sell, S.; Beutner, S.; Mayer, B.; Walsh, R. Chemistry of carotenoid oxidation and free radical reactions. *Pure Appl. Chem.* **1999**, *71*, 2253–2262. [[CrossRef](#)]
34. Kuroki, T.; Ikeda, S.; Okada, T.; Maoka, T.; Kitamura, A.; Sugimoto, M.; Kume, S. Astaxanthin ameliorates heat stress-induced impairment of blastocyst development In Vitro: Astaxanthin colocalization with and action on mitochondria. *J. Assist. Reprod. Genet.* **2013**, *30*, 623–631. [[CrossRef](#)] [[PubMed](#)]
35. Park, J.S.; Mathison, B.D.; Hayek, M.G.; Zhang, J.; Reinhart, G.A.; Chew, B.P. Astaxanthin modulates age-associated mitochondrial dysfunction in healthy dogs1. *J. Anim. Sci.* **2013**, *91*, 268–275. [[CrossRef](#)] [[PubMed](#)]
36. Ames, B.N. Dietary carcinogens and anticarcinogens. Oxygen radicals and degenerative diseases. *Science* **1983**, *221*, 1256–1264. [[CrossRef](#)] [[PubMed](#)]
37. Dušinská, M.; Kažimírová, A.; Barančoková, M.; Beňo, M.; Smolkova, B.; Horská, A.; Rašlová, K.; Wsólóvá, L.; Collins, A. Nutritional supplementation with antioxidants decreases chromosomal damage in humans. *Mutagenesis* **2003**, *18*, 371–376. [[CrossRef](#)]

38. Federico, A.; Morgillo, F.; Tuccillo, C.; Ciardiello, F.; Loguercio, C. Chronic inflammation and oxidative stress in human carcinogenesis. *Int. J. Cancer* **2007**, *121*, 2381–2386. [[CrossRef](#)]
39. Choi, H.D.; Youn, Y.K.; Shin, W.G. Positive Effects of Astaxanthin on Lipid Profiles and Oxidative Stress in Overweight Subjects. *Plant Foods Hum. Nutr.* **2011**, *66*, 363–369. [[CrossRef](#)]
40. Wolf, A.M.; Asoh, S.; Hiranuma, H.; Ohsawa, I.; Iio, K.; Satou, A.; Ishikura, M.; Ohta, S. Astaxanthin protects mitochondrial redox state and functional integrity against oxidative stress. *J. Nutr. Biochem.* **2010**, *21*, 381–389. [[CrossRef](#)]
41. Aoi, W.; Naito, Y.; Takamami, Y.; Ishii, T.; Kawai, Y.; Akagiri, S.; Kato, Y.; Osawa, T.; Yoshikawa, T. Astaxanthin improves muscle lipid metabolism in exercise via inhibitory effect of oxidative CPT I modification. *Biochem. Biophys. Res. Commun.* **2008**, *366*, 892–897. [[CrossRef](#)]
42. Park, J.S.; Chyun, J.H.; Kim, Y.K.; Line, L.L.; Chew, B.P. Astaxanthin decreased oxidative stress and inflammation and enhanced immune response in humans. *Nutr. Metab.* **2010**, *7*, 18. [[CrossRef](#)]
43. Kavitha, K.; Kowshik, J.; Kishore, T.K.K.; Baba, A.B.; Nagini, S. Astaxanthin inhibits NF- $\kappa$ B and Wnt/ $\beta$ -catenin signaling pathways via inactivation of Erk/MAPK and PI3K/Akt to induce intrinsic apoptosis in a hamster model of oral cancer. *Biochim. Biophys. Acta (BBA) Gen. Subj.* **2013**, *1830*, 4433–4444. [[CrossRef](#)]
44. Kowshik, J.; Baba, A.B.; Giri, H.; Reddy, G.D.; Dixit, M.; Nagini, S. Astaxanthin Inhibits JAK/STAT-3 Signaling to Abrogate Cell Proliferation, Invasion and Angiogenesis in a Hamster Model of Oral Cancer. *PLoS ONE* **2014**, *9*, e109114. [[CrossRef](#)] [[PubMed](#)]
45. Palozza, P.; Torelli, C.; Boninsegna, A.; Simone, R.; Catalano, A.; Mele, M.C.; Picci, N. Growth-inhibitory effects of the astaxanthin-rich alga *Haematococcus pluvialis* in human colon cancer cells. *Cancer Lett.* **2009**, *283*, 108–117. [[CrossRef](#)] [[PubMed](#)]
46. Tanaka, T.; Makita, H.; Ohnishi, M.; Mori, H.; Satoh, K.; Hara, A. Chemoprevention of rat oral carcinogenesis by naturally occurring xanthophylls, astaxanthin and canthaxanthin. *Cancer Res.* **1995**, *55*, 4059–4064. [[PubMed](#)]
47. Brito, A.; Ribeiro, M.; Abrantes, A.M.; Pires, A.; Teixeira, R.; Tralhão, J.; Botelho, M.F. Quercetin in Cancer Treatment, Alone or in Combination with Conventional Therapeutics? *Curr. Med. Chem.* **2015**, *22*, 3025–3039. [[CrossRef](#)]
48. Srivastava, S.; Somasagara, R.R.; Hegde, M.; Nishana, M.; Tadi, S.K.; Srivastava, M.; Choudhary, B.; Raghavan, S.C. Quercetin, a Natural Flavonoid Interacts with DNA, Arrests Cell Cycle and Causes Tumor Regression by Activating Mitochondrial Pathway of Apoptosis. *Sci. Rep.* **2016**, *6*, 24049. [[CrossRef](#)]
49. Satyan, K.; Swamy, N.; Dizon, D.S.; Singh, R.; Granai, C.O.; Brard, L. Phenethyl isothiocyanate (PEITC) inhibits growth of ovarian cancer cells by inducing apoptosis: Role of caspase and MAPK activation. *Gynecol. Oncol.* **2006**, *103*, 261–270. [[CrossRef](#)]
50. Wattenberg, L.W. Inhibitory effects of benzyl isothiocyanate administered shortly before diethylnitrosamine or benzo[a]pyrene on pulmonary and forestomach neoplasia in A/J mice. *Carcinogenesis* **1987**, *8*, 1971–1973. [[CrossRef](#)]
51. Long, Y.; Fei, H.; Xu, S.; Wen, J.; Ye, L.; Su, Z. Association about dietary vitamin C intake on the risk of ovarian cancer: A meta-analysis. *Biosci. Rep.* **2020**, *40*. [[CrossRef](#)]
52. L'Espérance, K.; Datta, G.D.; Qureshi, S.; Koushik, A. Vitamin D Exposure and Ovarian Cancer Risk and Prognosis. *Int. J. Environ. Res. Public Health* **2020**, *17*, 1168. [[CrossRef](#)]
53. Leng, Y.; Zhou, H.; Meng, F.; Tian, T.; Xu, J.; Yan, F. Association of vitamin E on the risk of ovarian cancer: A meta-analysis. *Biosci. Rep.* **2019**, *39*. [[CrossRef](#)]
54. Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group. The Effect of Vitamin E and Beta Carotene on the Incidence of Lung Cancer and Other Cancers in Male Smokers. *N. Engl. J. Med.* **1994**, *330*, 1029–1035. [[CrossRef](#)]
55. Wang, Q.; He, C. Dietary vitamin A intake and the risk of ovarian cancer: A meta-analysis. *Biosci. Rep.* **2020**, *40*, 40. [[CrossRef](#)] [[PubMed](#)]
56. Hua, X.; Yu, L.; You, R.; Yang, Y.; Liao, J.; Chen, D.; Yu, L. Association among Dietary Flavonoids, Flavonoid Subclasses and Ovarian Cancer Risk: A Meta-Analysis. *PLoS ONE* **2016**, *11*, e0151134. [[CrossRef](#)] [[PubMed](#)]
57. Chang, E.T.; Lee, V.S.; Canchola, A.J.; Clarke, C.A.; Purdie, D.M.; Reynolds, P.; Anton-Culver, H.; Bernstein, L.; Deapen, D.; Peel, D.; et al. Diet and Risk of Ovarian Cancer in the California Teachers Study Cohort. *Am. J. Epidemiol.* **2007**, *165*, 802–813. [[CrossRef](#)]
58. Warburg, O. On the Origin of Cancer Cells. *Science* **1956**, *123*, 309–314. [[CrossRef](#)] [[PubMed](#)]
59. Kato, N.; Tamura, G.; Motoyama, T. Hypomethylation of hepatocyte nuclear factor-1beta (HNF-1beta) CpG island in clear cell carcinoma of the ovary. *Virchows Archiv.* **2008**, *452*, 175–180. [[CrossRef](#)]
60. Zhu, J.-N.; Jiang, L.; Jiang, J.-H.; Yang, X.; Li, X.-Y.; Zeng, J.-X.; Shi, R.-Y.; Shi, Y.; Pan, X.-R.; Han, Z.-P.; et al. Hepatocyte nuclear factor-1beta enhances the stemness of hepatocellular carcinoma cells through activation of the Notch pathway. *Sci. Rep.* **2017**, *7*, 4793. [[CrossRef](#)]
61. Sugiyama, T.; Kamura, T.; Kigawa, J.; Terakawa, N.; Kikuchi, Y.; Kita, T.; Suzuki, M.; Sato, I.; Taguchi, K. Clinical characteristics of clear cell carcinoma of the ovary: A distinct histologic type with poor prognosis and resistance to platinum-based chemotherapy. *Cancer* **2000**, *88*, 2584–2589. [[CrossRef](#)]
62. Goff, B.A.; De La Cuesta, R.S.; Muntz, H.G.; Fleischhacker, D.; Ek, M.; Rice, L.W.; Nikrui, N.; Tamimi, H.K.; Cain, J.M.; Greer, B.E.; et al. Clear Cell Carcinoma of the Ovary: A Distinct Histologic Type with Poor Prognosis and Resistance to Platinum-Based Chemotherapy in Stage III Disease. *Gynecol. Oncol.* **1996**, *60*, 412–417. [[CrossRef](#)]
63. Senkel, S.; Lucas, B.; Klein-Hitpass, L.; Ryyfel, G.U. Identification of target genes of the transcription factor HNF1 $\beta$  and HNF1 $\alpha$  in a human embryonic kidney cell line. *Biochim. Biophys. Acta (BBA) Gene Struct. Expr.* **2005**, *1731*, 179–190. [[CrossRef](#)]

64. Okamoto, T.; Mandai, M.; Matsumura, N.; Yamaguchi, K.; Kondoh, H.; Amano, Y.; Baba, T.; Hamanishi, J.; Abiko, K.; Kosaka, K.; et al. Hepatocyte nuclear factor-1 $\beta$  (HNF-1 $\beta$ ) promotes glucose uptake and glycolytic activity in ovarian clear cell carcinoma. *Mol. Carcinog.* **2015**, *54*, 35–49. [CrossRef] [PubMed]
65. Liu, P.; Khurana, A.; Rattan, R.; He, X.; Kalloger, S.; Dowdy, S.; Gilks, B.; Shridhar, V. Regulation of HSulf-1 Expression by Variant Hepatic Nuclear Factor 1 in Ovarian Cancer. *Cancer Res.* **2009**, *69*, 4843–4850. [CrossRef] [PubMed]
66. Chen, C.; Zhou, Y.; Hu, C.; Wang, Y.; Yan, Z.; Li, Z.; Wu, R. Mitochondria and oxidative stress in ovarian endometriosis. *Free Radic. Biol. Med.* **2019**, *136*, 22–34. [CrossRef] [PubMed]
67. Hemachandra, L.P.M.P.; Shin, D.-H.; Dier, U.; Iuliano, J.N.; Engelberth, S.A.; Uusitalo, L.M.; Murphy, S.K.; Hempel, N. Mitochondrial Superoxide Dismutase Has a Protumorigenic Role in Ovarian Clear Cell Carcinoma. *Cancer Res.* **2015**, *75*, 4973–4984. [CrossRef] [PubMed]
68. Amano, T.; Chano, T.; Isono, T.; Kimura, F.; Kushima, R.; Murakami, T. Abundance of mitochondrial superoxide dismutase is a negative predictive biomarker for endometriosis-associated ovarian cancers. *World J. Surg. Oncol.* **2019**, *17*, 1–7. [CrossRef]
69. Mizuno, T.; Suzuki, N.; Makino, H.; Furui, T.; Morii, E.; Aoki, H.; Kunisada, T.; Yano, M.; Kuji, S.; Hirashima, Y.; et al. Cancer stem-like cells of ovarian clear cell carcinoma are enriched in the ALDH-high population associated with an accelerated scavenging system in reactive oxygen species. *Gynecol. Oncol.* **2015**, *137*, 299–305. [CrossRef]
70. Visvader, J.E.; Lindeman, G.J. Cancer stem cells in solid tumours: Accumulating evidence and unresolved questions. *Nat. Rev. Cancer* **2008**, *8*, 755–768. [CrossRef]
71. Chano, T.; Kita, H.; Avnet, S.; Lemma, S.; Baldini, N. Prominent role of RAB39A-RXR $\beta$  axis in cancer development and stemness. *Oncotarget* **2018**, *9*, 9852–9866. [CrossRef]
72. Metzler, M.A.; Sandell, L.L. Enzymatic Metabolism of Vitamin A in Developing Vertebrate Embryos. *Nutrients* **2016**, *8*, 812. [CrossRef]
73. Shannon, S.R.; Moise, A.R.; Trainor, P. New insights and changing paradigms in the regulation of vitamin A metabolism in development. *Wiley Interdiscip. Rev. Dev. Biol.* **2017**, *6*, e264. [CrossRef]
74. Obrochta, K.M.; Krois, C.R.; Campos, B.; Napoli, J.L.; Duan, X.-L.; Liu, N.-N.; Yang, Y.-T.; Li, H.-H.; Li, M.; Dou, S.-X.; et al. Insulin Regulates Retinol Dehydrogenase Expression and All-trans-retinoic Acid Biosynthesis through FoxO1. *J. Biol. Chem.* **2015**, *290*, 7259–7268. [CrossRef] [PubMed]
75. Felix, E.L.; Loyd, B.; Cohen, M.H. Inhibition of the growth and development of a transplantable murine melanoma by vitamin A. *Science* **1975**, *189*, 886–888. [CrossRef] [PubMed]
76. Dillehay, D.L.; Shealy, Y.F.; Lamon, E.W. Inhibition of Moloney murine lymphoma and sarcoma growth in vivo by dietary retinoids. *Cancer Res.* **1989**, *49*, 44–50. [PubMed]
77. Aebi, S.; Kröning, R.; Cenni, B.; Sharma, A.; Fink, D.; Los, G.; Weisman, R.; Howell, S.B.; Christen, R.D. all-trans retinoic acid enhances cisplatin-induced apoptosis in human ovarian adenocarcinoma and in squamous head and neck cancer cells. *Clin. Cancer Res.* **1997**, *3*, 2033–2038.
78. Schug, T.T.; Berry, D.C.; Shaw, N.S.; Travis, S.N.; Noy, N. Opposing Effects of Retinoic Acid on Cell Growth Result from Alternate Activation of Two Different Nuclear Receptors. *Cell* **2007**, *129*, 723–733. [CrossRef]
79. Available online: <https://www.proteinatlas.org/ENSG00000164687-FABP5/pathology> (accessed on 15 December 2020).
80. Guan, F.; Wang, L.; Hao, S.; Wu, Z.; Bai, J.; Kang, Z.; Zhou, Q.; Chang, H.; Yin, H.; Li, D.; et al. Retinol dehydrogenase-10 promotes development and progression of human glioma via the TWEAK-NF- $\kappa$ B axis. *Oncotarget* **2017**, *8*, 105262–105275. [CrossRef]
81. Fujimura, M.; Katsumata, N.; Tsuda, H.; Uchi, N.; Miyazaki, S.; Hidaka, T.; Sakai, M.; Saito, S. HER2 Is Frequently Over-expressed in Ovarian Clear Cell Adenocarcinoma: Possible Novel Treatment Modality Using Recombinant Monoclonal Antibody against HER2, Trastuzumab. *Jpn. J. Cancer Res.* **2002**, *93*, 1250–1257. [CrossRef]
82. Kim, H.-J.; Yoon, A.; Ryu, J.-Y.; Cho, Y.-J.; Choi, J.-J.; Song, S.Y.; Bang, H.; Lee, J.S.; Cho, W.C.; Choi, C.H.; et al. c-MET as a Potential Therapeutic Target in Ovarian Clear Cell Carcinoma. *Sci. Rep.* **2016**, *6*, 38502. [CrossRef]
83. Wakinoue, S.; Chano, T.; Amano, T.; Isono, T.; Kimura, F.; Kushima, R.; Murakami, T. ADP-ribosylation factor-like 4C predicts worse prognosis in endometriosis-associated ovarian cancers. *Cancer Biomark.* **2019**, *24*, 223–229. [CrossRef]
84. Konstantinopoulos, P.A.; Brady, W.E.; Farley, J.H.; Armstrong, A.; Uyar, D.; Gershenson, D. Phase II study of single-agent cabozantinib in patients with recurrent clear cell ovarian, primary peritoneal or fallopian tube cancer (NRG-GY001). *Gynecol. Oncol.* **2018**, *150*, 9–13. [CrossRef]
85. Farley, J.H.; Brady, W.E.; Fujiwara, K.; Nomura, H.; Yunokawa, M.; Tokunaga, H.; Saitou, M.; Gershenson, D.M. A phase II evaluation of temsirolimus in combination with carboplatin and paclitaxel followed by temsirolimus consolidation as first-line therapy in the treatment of stage III-IV clear cell carcinoma of the ovary. *J. Clin. Oncol.* **2016**, *34*, 5531. [CrossRef]
86. DeVorkin, L.; Hattersley, M.; Kim, P.; Ries, J.; Spowart, J.; Anglesio, M.S.; Levi, S.M.; Huntsman, D.G.; Amaravadi, R.K.; Winkler, J.D.; et al. Autophagy Inhibition Enhances Sunitinib Efficacy in Clear Cell Ovarian Carcinoma. *Mol. Cancer Res.* **2017**, *15*, 250–258. [CrossRef] [PubMed]
87. Bitler, B.G.; Aird, K.M.; Garipov, A.; Li, H.; Amatangelo, M.; Kossenkov, A.V.; Schultz, D.C.; Liu, Q.; Shih, I.-M.; Conejo-Garcia, J.R.; et al. Synthetic lethality by targeting EZH2 methyltransferase activity in ARID1A-mutated cancers. *Nat. Med.* **2015**, *21*, 231–238. [CrossRef] [PubMed]

88. Berns, K.; Caumanns, J.J.; Hijmans, E.M.; Gennissen, A.M.C.; Severson, T.M.; Evers, B.; Wisman, G.B.A.; Meersma, G.J.; Lieftink, C.; Beijersbergen, R.L.; et al. ARID1A mutation sensitizes most ovarian clear cell carcinomas to BET inhibitors. *Oncogene* **2018**, *37*, 4611–4625. [[CrossRef](#)] [[PubMed](#)]
89. Ogiwara, H.; Takahashi, K.; Sasaki, M.; Kuroda, T.; Yoshida, H.; Watanabe, R.; Maruyama, A.; Makinoshima, H.; Chiwaki, F.; Sasaki, H.; et al. Targeting the Vulnerability of Glutathione Metabolism in ARID1A-Deficient Cancers. *Cancer Cell* **2019**, *35*, 177–190.e8. [[CrossRef](#)]
90. Wiegand, K.C.; Hennessy, B.T.; Leung, S.; Wang, Y.; Ju, Z.; Murray, M.; Kalloger, S.; Finlayson, S.; Stemke-Hale, K.; Lu, Y.; et al. A functional proteogenomic analysis of endometrioid and clear cell carcinomas using reverse phase protein array and mutation analysis: Protein expression is histotype-specific and loss of ARID1A/BAF250a is associated with AKT phosphorylation. *BMC Cancer* **2014**, *14*, 120. [[CrossRef](#)]
91. Le, D.T.; Durham, J.N.; Smith, K.N.; Wang, H.; Bartlett, B.R.; Aulakh, L.K.; Lu, S.; Kemberling, H.; Wilt, C.; Lubner, B.S.; et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science* **2017**, *357*, 409–413. [[CrossRef](#)]
92. Le, D.T.; Uram, J.N.; Wang, H.; Bartlett, B.R.; Kemberling, H.; Eyring, A.D.; Skora, A.D.; Lubner, B.S.; Azad, N.S.; Laheru, D.; et al. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. *N. Engl. J. Med.* **2015**, *372*, 2509–2520. [[CrossRef](#)]
93. Samstein, R.M.; Lee, C.-H.; Shoushtari, A.N.; Hellmann, M.D.; Shen, R.; Janjigian, Y.Y.; Barron, D.A.; Zehir, A.; Jordan, E.J.; Omuro, A.; et al. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. *Nat. Genet.* **2019**, *51*, 202–206. [[CrossRef](#)]
94. Yarchoan, M.; Hopkins, A.; Jaffee, E.M. Tumor Mutational Burden and Response Rate to PD-1 Inhibition. *N. Engl. J. Med.* **2017**, *377*, 2500–2501. [[CrossRef](#)]
95. Akbari, M.R.; Zhang, S.; Cragun, D.; Lee, J.-H.; Coppola, D.; McLaughlin, J.R.; Risch, H.A.; Rosen, B.; Shaw, P.; Sellers, T.A.; et al. Correlation between germline mutations in MMR genes and microsatellite instability in ovarian cancer specimens. *Fam. Cancer* **2017**, *16*, 351–355. [[CrossRef](#)] [[PubMed](#)]
96. Feinberg, J.; Elvin, J.A.; Bellone, S.; Santin, A.D. Identification of ovarian cancer patients for immunotherapy by concurrent assessment of tumor mutation burden (TMB), microsatellite instability (MSI) status, and targetable genomic alterations (GA). *Gynecol. Oncol.* **2018**, *149* (Suppl. 1), 36. [[CrossRef](#)]
97. Jiang, T.; Chen, X.; Su, C.; Ren, S.; Zhou, C. Pan-cancer analysis of ARID1A Alterations as Biomarkers for Immunotherapy Outcomes. *J. Cancer* **2020**, *11*, 776–780. [[CrossRef](#)] [[PubMed](#)]
98. Thomas, L.J.; Vitale, L.; O'Neill, T.; Dolnick, R.Y.; Wallace, P.K.; Minderman, H.; Gergel, L.E.; Forsberg, E.M.; Boyer, J.M.; Storey, J.R.; et al. Development of a Novel Antibody-Drug Conjugate for the Potential Treatment of Ovarian, Lung, and Renal Cell Carcinoma Expressing TIM-1. *Mol. Cancer Ther.* **2016**, *15*, 2946–2954. [[CrossRef](#)] [[PubMed](#)]
99. Goto, M.; Miwa, H.; Shikami, M.; Tsunekawa-Imai, N.; Suganuma, K.; Mizuno, S.; Takahashi, M.; Mizutani, M.; Hanamura, I.; Nitta, M. Importance of Glutamine Metabolism in Leukemia Cells by Energy Production Through TCA Cycle and by Redox Homeostasis. *Cancer Investig.* **2014**, *32*, 241–247. [[CrossRef](#)] [[PubMed](#)]
100. Roesch, A.; Vultur, A.; Bogeski, I.; Wang, H.; Zimmermann, K.M.; Speicher, D.W.; Körbel, C.; Laschke, M.W.; Gimotty, P.A.; Philipp, S.E.; et al. Overcoming Intrinsic Multidrug Resistance in Melanoma by Blocking the Mitochondrial Respiratory Chain of Slow-Cycling JARID1Bhigh Cells. *Cancer Cell* **2013**, *23*, 811–825. [[CrossRef](#)] [[PubMed](#)]
101. Vazquez, F.; Lim, J.-H.; Chim, H.; Bhalla, K.; Girmun, G.; Pierce, K.; Clish, C.B.; Granter, S.R.; Widlund, H.R.; Spiegelman, B.M.; et al. PGC1 $\alpha$  Expression Defines a Subset of Human Melanoma Tumors with Increased Mitochondrial Capacity and Resistance to Oxidative Stress. *Cancer Cell* **2013**, *23*, 287–301. [[CrossRef](#)]
102. Viale, A.; Pettazoni, P.; Lyssiotis, C.A.; Ying, H.; Sanchez, N.; Marchesini, M.; Carugo, A.; Green, T.; Seth, S.; Giuliani, V.; et al. Oncogene ablation-resistant pancreatic cancer cells depend on mitochondrial function. *Nature* **2014**, *514*, 628–632. [[CrossRef](#)]
103. Molina, J.R.; Sun, Y.; Protopopova, M.; Gera, S.; Bandi, M.; Bristow, C.; McAfoos, T.; Morlacchi, P.; Ackroyd, J.J.; Agip, A.-N.A.; et al. An inhibitor of oxidative phosphorylation exploits cancer vulnerability. *Nat. Med.* **2018**, *24*, 1036–1046. [[CrossRef](#)]
104. El-Mir, M.-Y.; Nogueira, V.; Fontaine, E.; Avéret, N.; Rigoulet, M.; Leverve, X. Dimethylbiguanide Inhibits Cell Respiration via an Indirect Effect Targeted on the Respiratory Chain Complex I. *J. Biol. Chem.* **2000**, *275*, 223–228. [[CrossRef](#)]
105. Thakur, S.; Daley, B.; Gaskins, K.; Vasko, V.V.; Boufraquech, M.; Patel, D.; Sourbier, C.; Reece, J.M.; Cheng, S.-Y.; Kebebew, E.; et al. Metformin Targets Mitochondrial Glycerophosphate Dehydrogenase to Control Rate of Oxidative Phosphorylation and Growth of Thyroid Cancer In Vitro and In Vivo. *Clin. Cancer Res.* **2018**, *24*, 4030–4043. [[CrossRef](#)] [[PubMed](#)]
106. Wheaton, W.W.; Weinberg, S.E.; Hamanaka, R.B.; Soberanes, S.; Sullivan, L.B.; Anso, E.; Glasauer, A.; Dufour, E.; Mutlu, G.M.; Budigner, G.S.; et al. Metformin inhibits mitochondrial complex I of cancer cells to reduce tumorigenesis. *eLife* **2014**, *3*, e02242. [[CrossRef](#)] [[PubMed](#)]
107. Bridges, H.R.; Jones, A.J.Y.; Pollak, M.N.; Hirst, J. Effects of metformin and other biguanides on oxidative phosphorylation in mitochondria. *Biochem. J.* **2014**, *462*, 475–487. [[CrossRef](#)] [[PubMed](#)]
108. Wilcock, C.; Bailey, C.J. Accumulation of metformin by tissues of the normal and diabetic mouse. *Xenobiotica* **1994**, *24*, 49–57. [[CrossRef](#)]
109. Owen, M.R.; Doran, E.; Halestrap, A.P. Evidence that metformin exerts its anti-diabetic effects through inhibition of complex 1 of the mitochondrial respiratory chain. *Biochem. J.* **2000**, *348*, 607–614. [[CrossRef](#)]
110. Eikawa, S.; Nishida, M.; Mizukami, S.; Yamazaki, C.; Nakayama, E.; Udono, H. Immune-mediated antitumor effect by type 2 diabetes drug, metformin. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 1809–1814. [[CrossRef](#)] [[PubMed](#)]

111. Sanada, S.; Futami, K.; Terada, A.; Yonemoto, K.; Ogasawara, S.; Akiba, J.; Yasumoto, M.; Sumi, A.; Ushijima, K.; Kamura, T.; et al. RECQL1 DNA Repair Helicase: A Potential Therapeutic Target and a Proliferative Marker against Ovarian Cancer. *PLoS ONE* **2013**, *8*, e72820. [[CrossRef](#)]
112. Arai, A.; Chano, T.; Futami, K.; Furuichi, Y.; Ikebuchi, K.; Inui, T.; Tameno, H.; Ochi, Y.; Shimada, T.; Hisa, Y.; et al. RECQL1 and WRN Proteins Are Potential Therapeutic Targets in Head and Neck Squamous Cell Carcinoma. *Cancer Res.* **2011**, *71*, 4598–4607. [[CrossRef](#)] [[PubMed](#)]
113. Chan, E.M.; Shibue, T.; McFarland, J.M.; Gaeta, B.; Ghandi, M.; Dumont, N.; Gonzalez, A.; McPartlan, J.S.; Li, T.; Zhang, Y.; et al. WRN helicase is a synthetic lethal target in microsatellite unstable cancers. *Nat. Cell Biol.* **2019**, *568*, 551–556. [[CrossRef](#)]
114. Sakharkar, M.K.; Sakharkar, K.R.; Pervaiz, S. Druggability of human disease genes. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 1156–1164. [[CrossRef](#)]